

5V569 - Atelier de allergie

Microbiota and Allergy

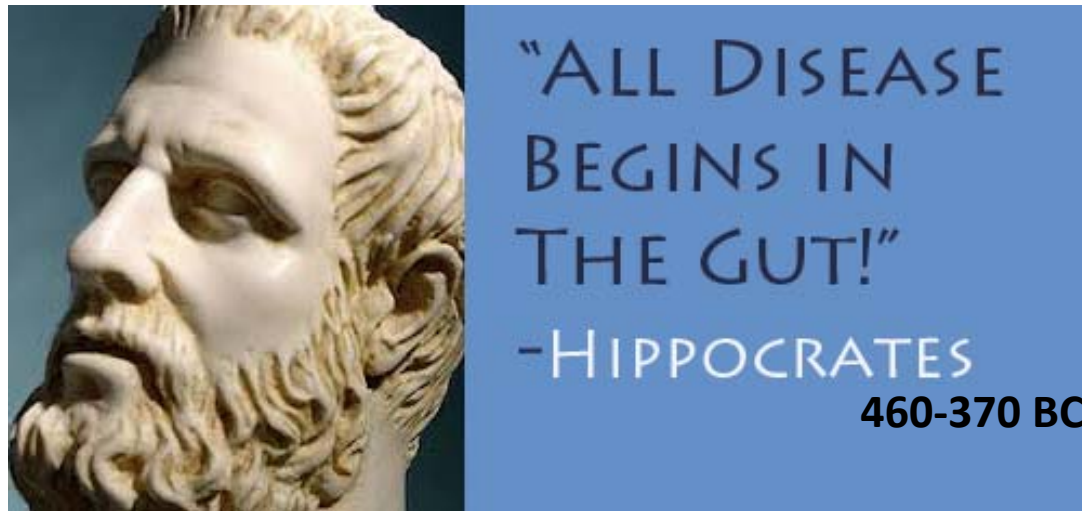
Martin LARSEN

INSERM U1135, CHU Pitié-Salpêtrière, Paris, France

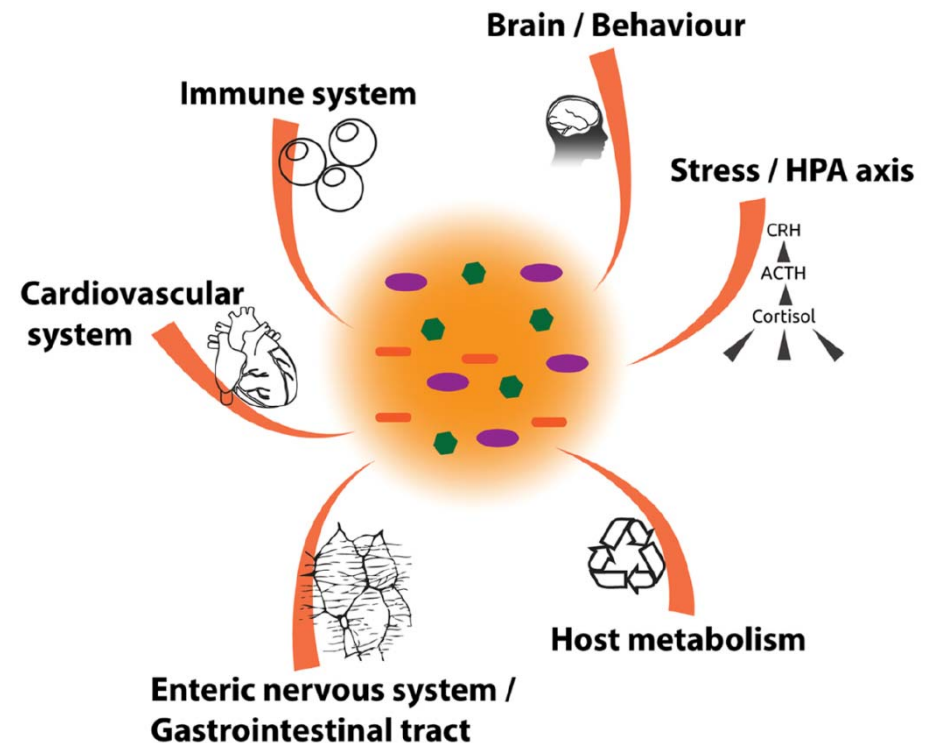
Outline

1. Model of allergy mechanisms
2. The gut microbiota and our digestive system
3. Gut microbiota and host immunity
4. Antibody responses
5. Gut microbiota and its role in disease
6. Gut microbiota in early life
7. Self-non-self versus the danger model.
8. Gut microbiota and allergy
9. Solutions

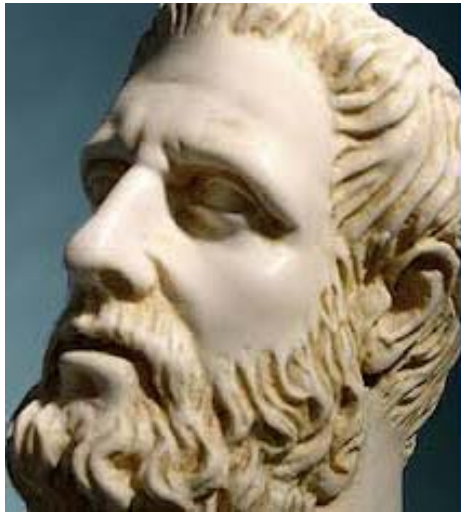
Why don't we all develop allergy? Search for the cause.



- First to be recognized for systematically using diet and exercise to treat life-style diseases.



Dysbiosis - cause or consequence - and so what?



"ALL DISEASE
BEGINS IN
THE GUT!"
-HIPPOCRATES
460-370 BC

"Le malade imaginaire"

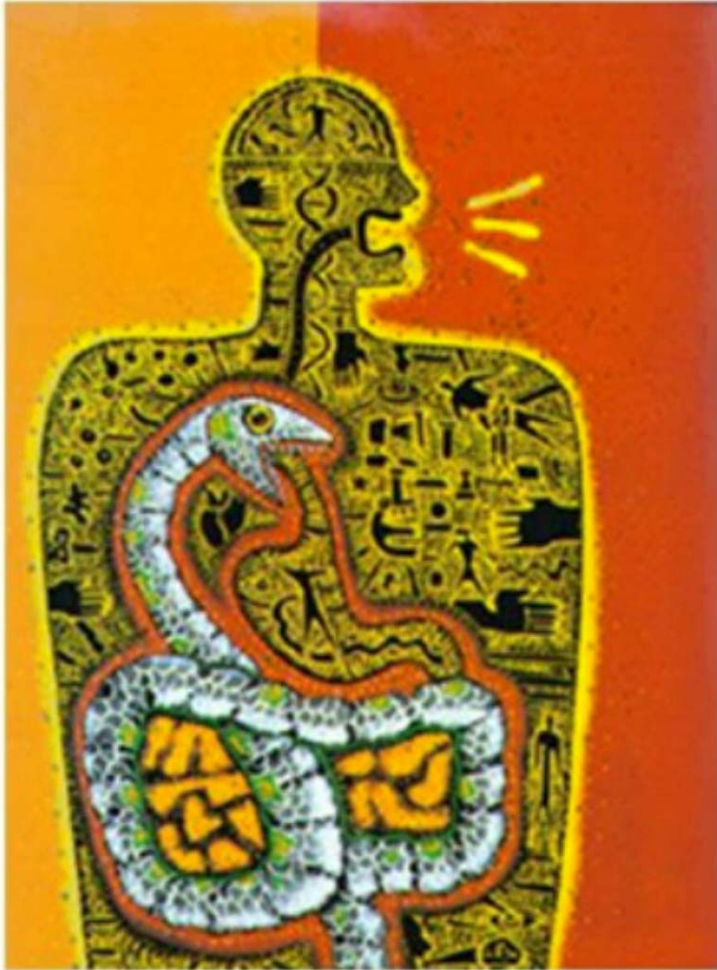
Molière, 1673



- Prescription of "lavement" for:
 - Digestion, intestinal secretion and bad mood.
- Dates back at least to 100 B.C.



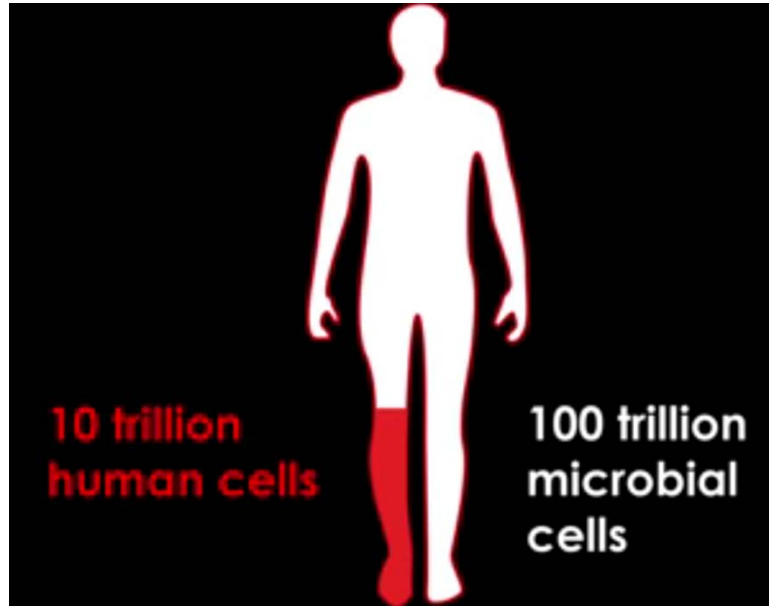
The human Gut and its inhabitants in numbers



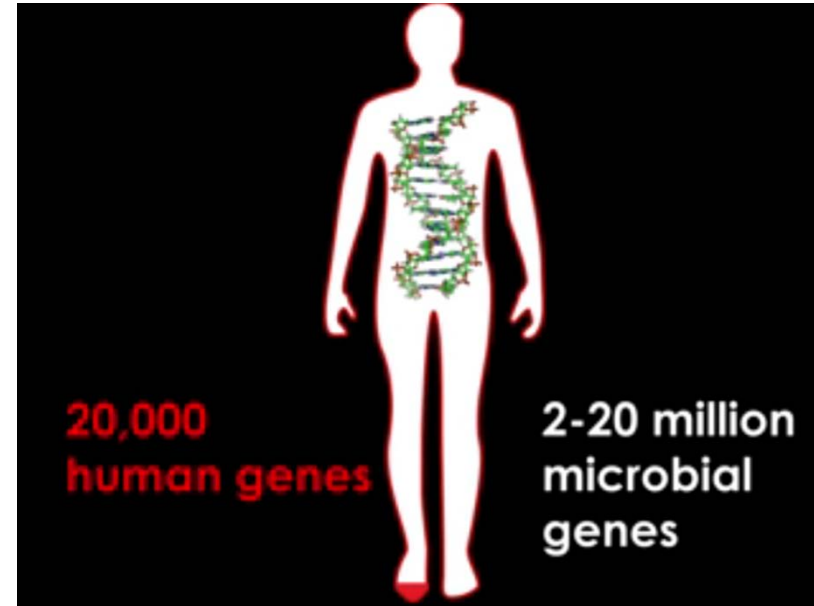
- 30 tons of food and 50.000 L during a lifetime
- Huge mucosal surface: 150-200 m²
- >50 billions of new bacteria every day
- 70-80% of all immune cells are located in the Gut.
- 1-2g secretory IgA per day
- 100 millions of neurons (as many as in the spinal cord).
- 10¹⁴ bacteria: x10 number of cells in the entire body, i.e. 1-2 kg.
- 100 times more bacterial genes than human genes.

We are outnumbered and outsmarted

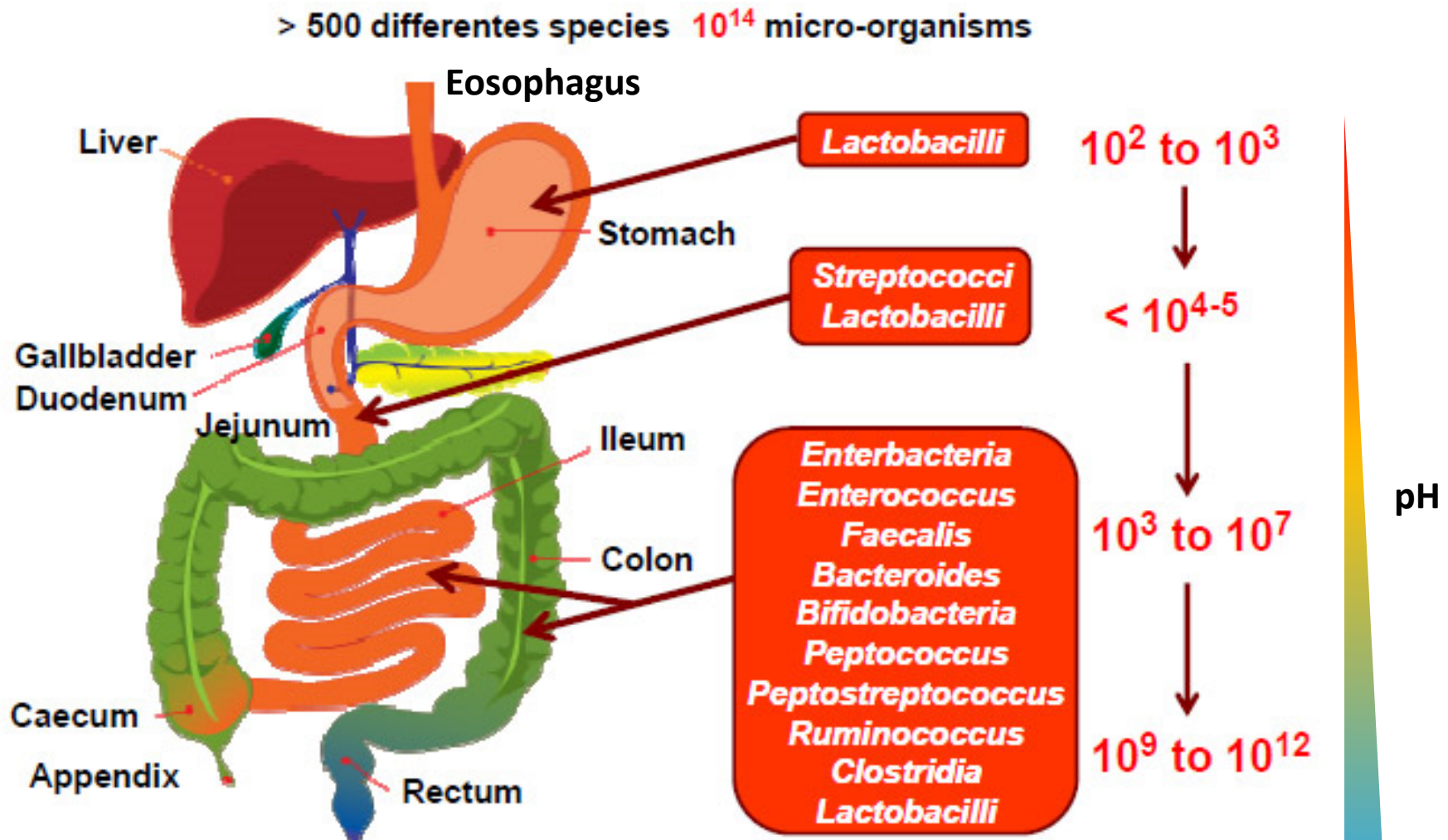
**Outnumbered
(1 to 10)**



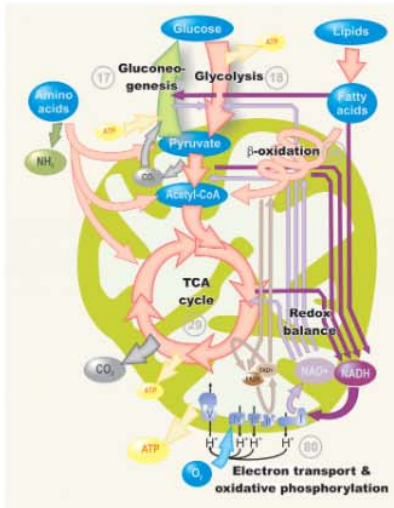
**Outsmarted
(100-1000 fold more genes)**



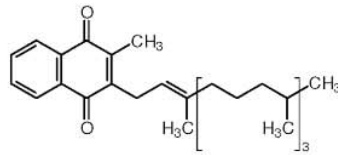
Spatial distribution of gut microbiota



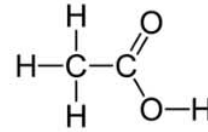
The many roles of gut Microbiota



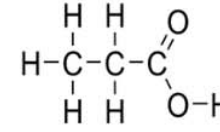
- **Increases the metabolic capacity of the host.**
 - **Digestion of otherwise unused food components.**



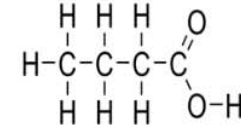
Vitamin synthesis
(eg Vitamin K)



Acetic acid (acetate)



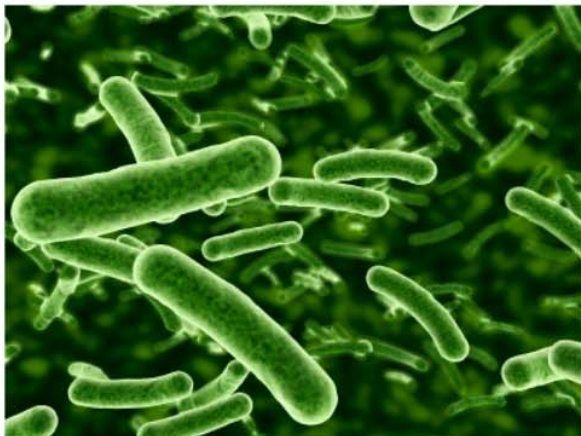
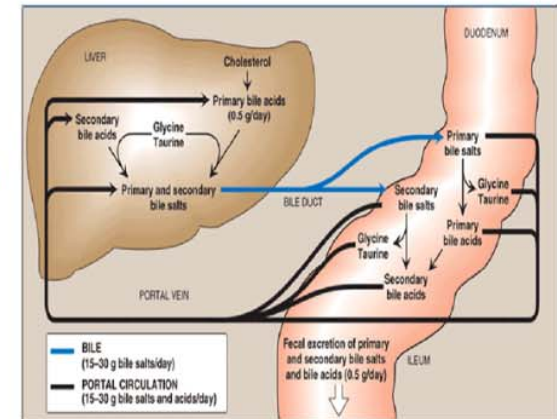
Propionic acid (propionate)



Butyric acid (butyrate)

Production of short chain fatty acids

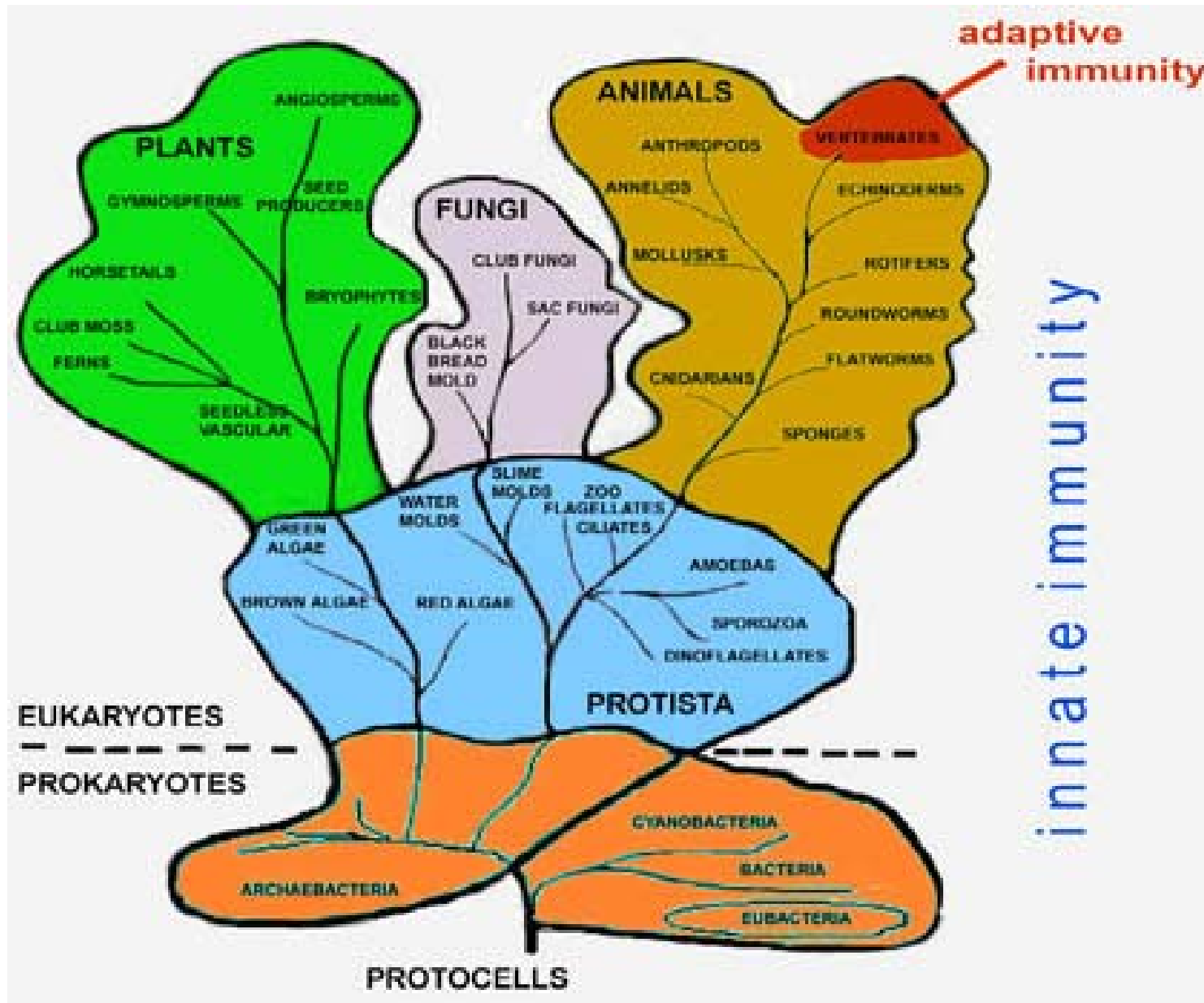
Completion of the bile-salt cycle Lipid uptake



Protect the host from colonization with pathogenic bacteria (Colonization resistance)

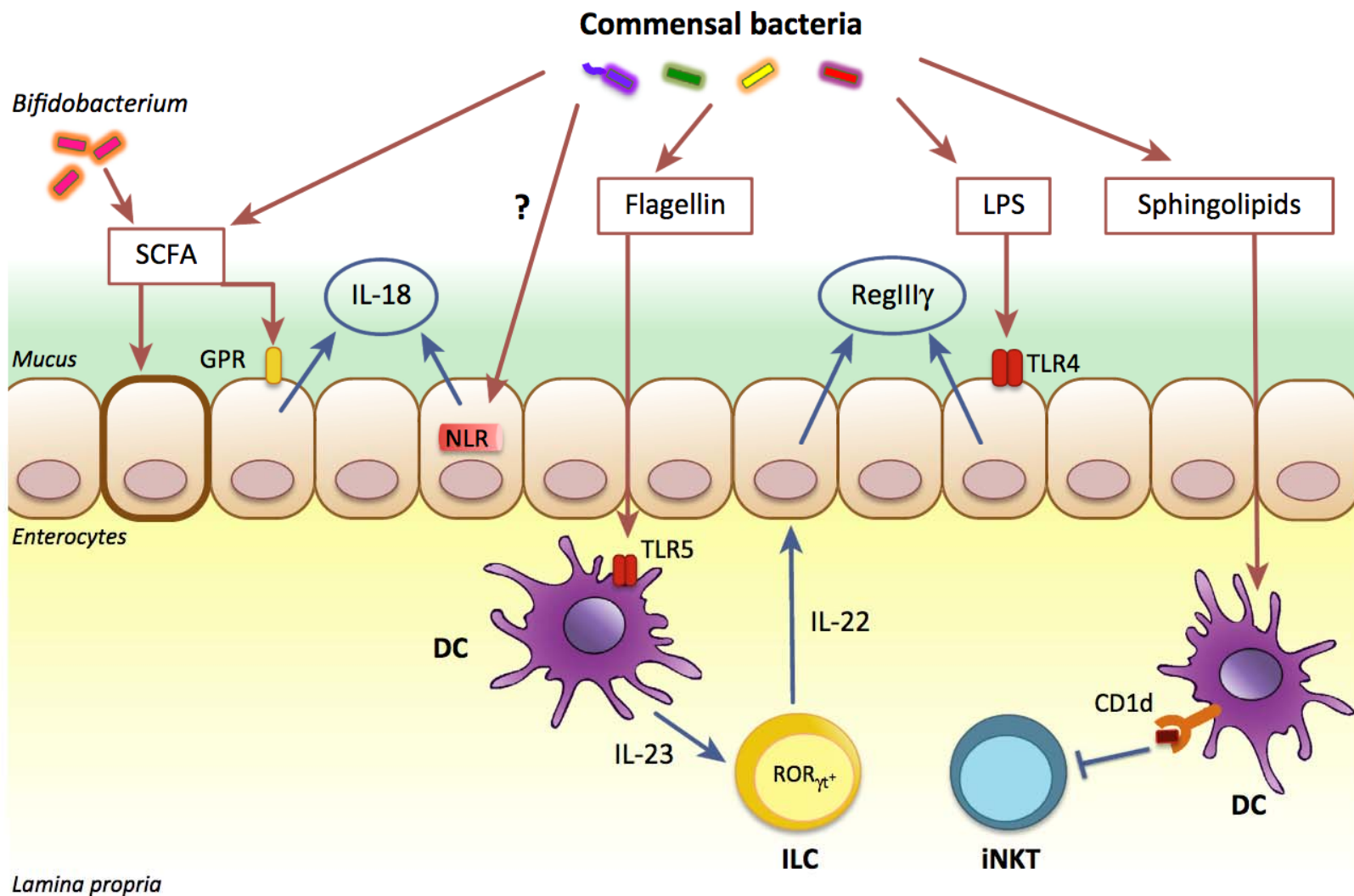
Immunity

Evolution of Adaptive immunity



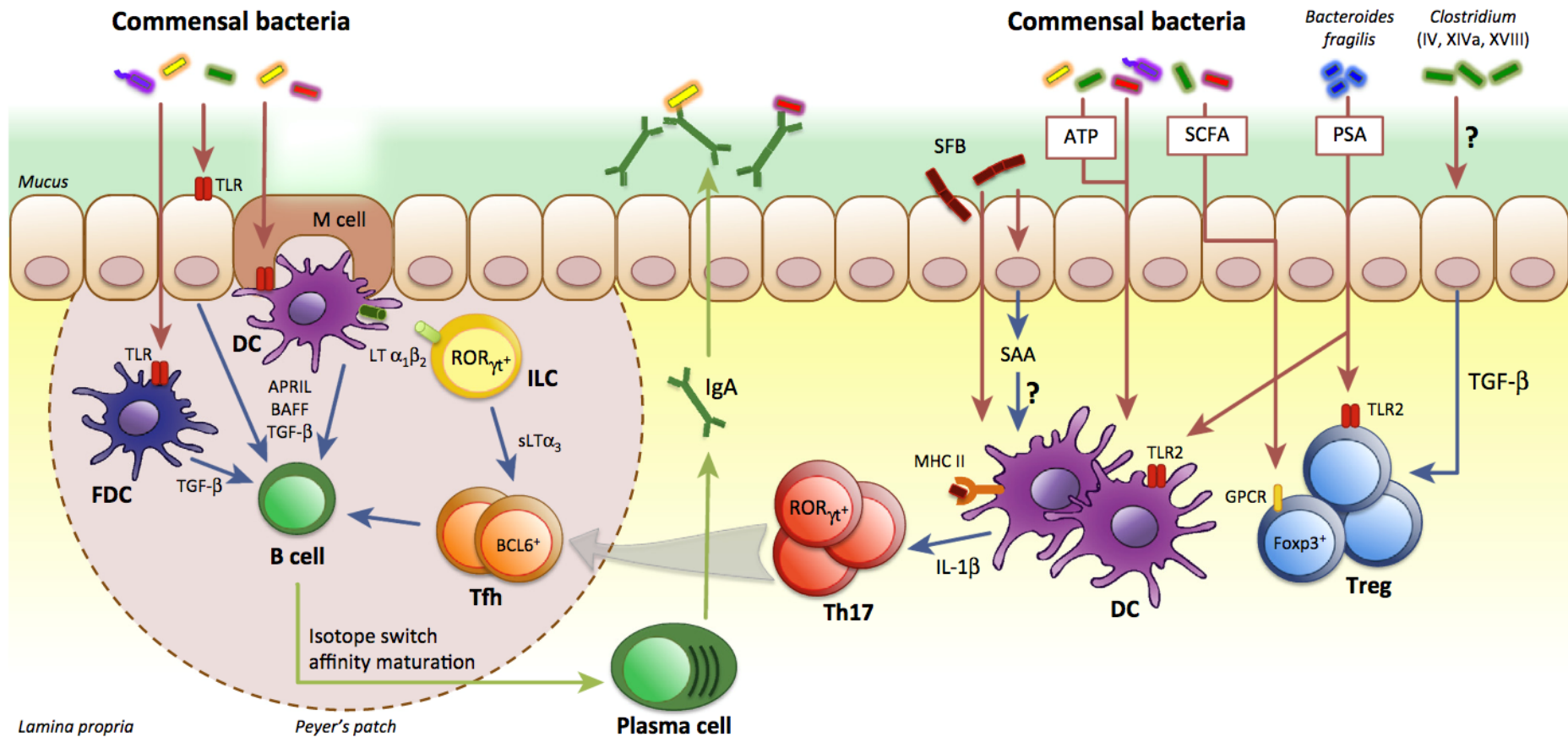
Gut Microbiota in health - Innate immunity

- Microbiota influences intestinal immune responses primarily through the expression of pathogen-associated molecular patterns (PAMPs) and metabolic by-products.



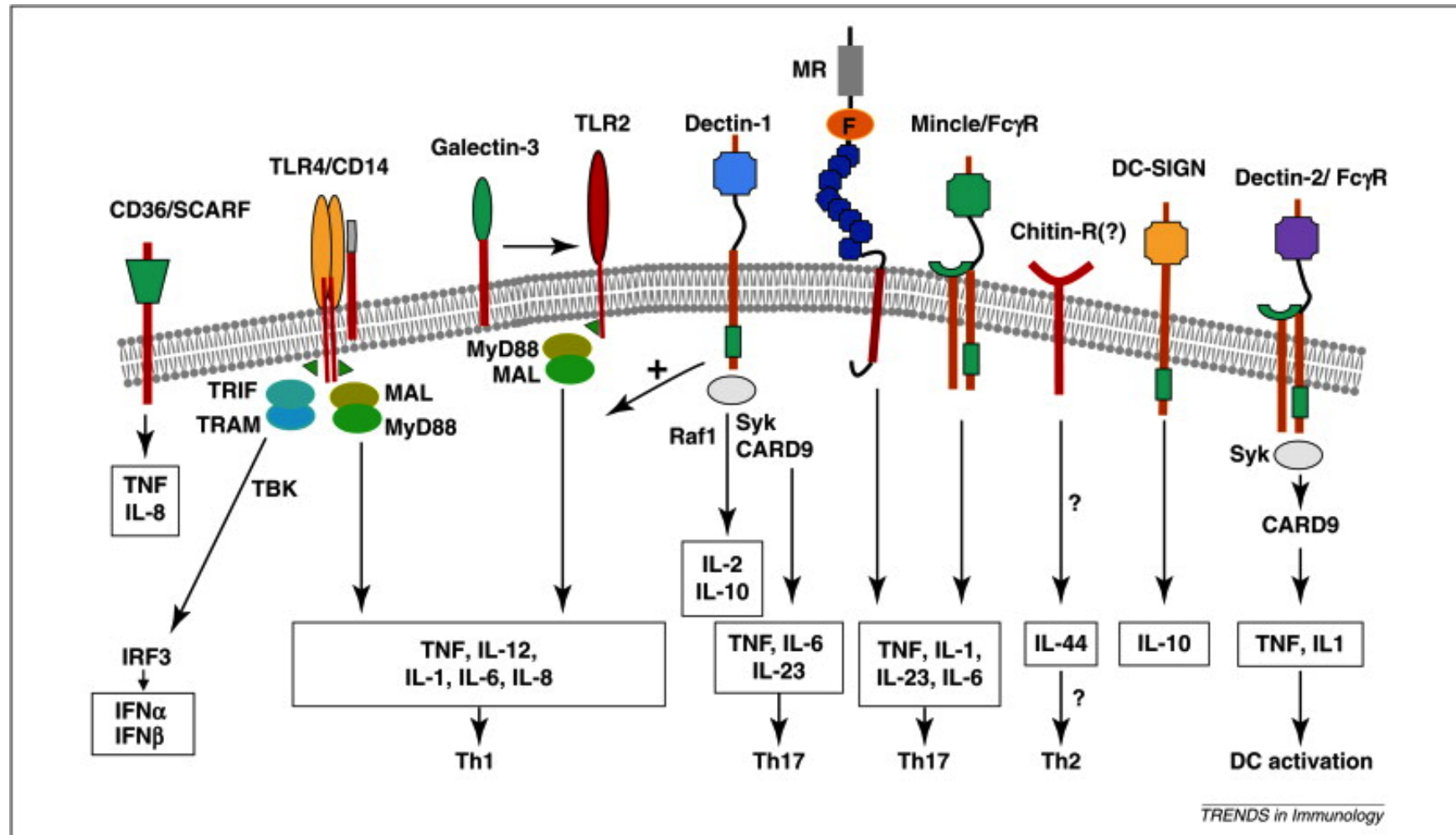
Gut Microbiota in health - adaptive immunity

- Microbiota stimulation leads to B cell switch to IgA, regulatory T cell induction, T cell differentiation to Th17



SAA=Serum amyloid A; GPCR=G protein-coupled receptor; PSA=Polysaccharide A; ILC=Innate lymphoid cells

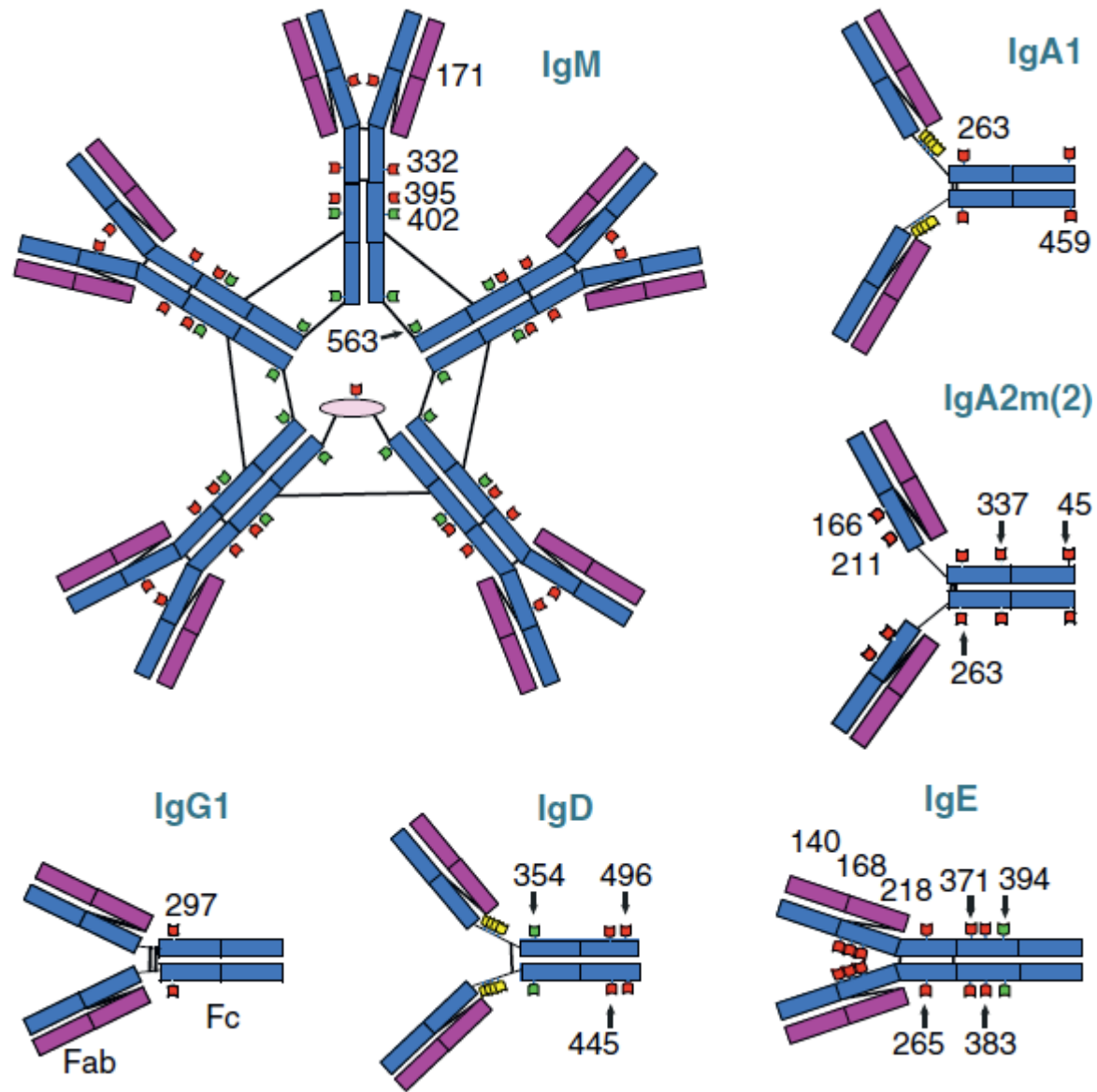
Microbiota, Toll-like receptors and T cell differentiation



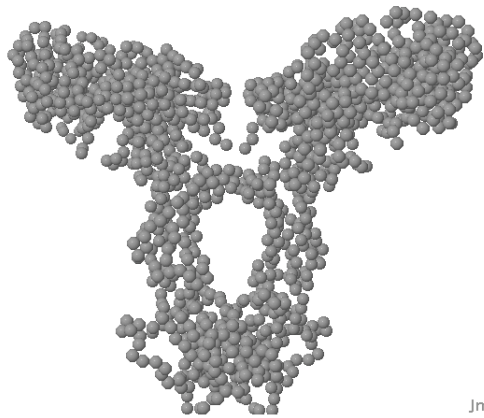
- House Dust Mite derived chitin induce IgE responses via a TLR2 dependent pathway (independent of TLR4 and 6)

Choi *et al.* All Asthma Immunol Res 2016

Antibody characteristics



Antibody characteristics

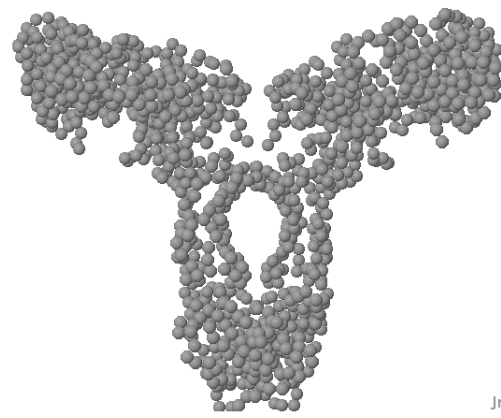


IgG

Small mammals
Higher monkeys

Y-shape
Sensitive to proteases

Non-Secretory



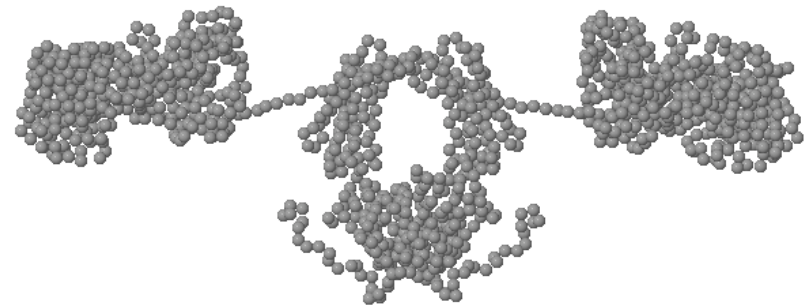
IgA2

Small mammals
Higher monkeys

Y-shape
Non-Sensitive to proteases

Secretory : non-planar (fab
fragment not aligned with Fc
portion)

Binds to Peyer's Patch M cells
to undergo transcytosis
thereby delivering antigens to
GALT
Mantis NJ *et al.* JI 2002



IgA1

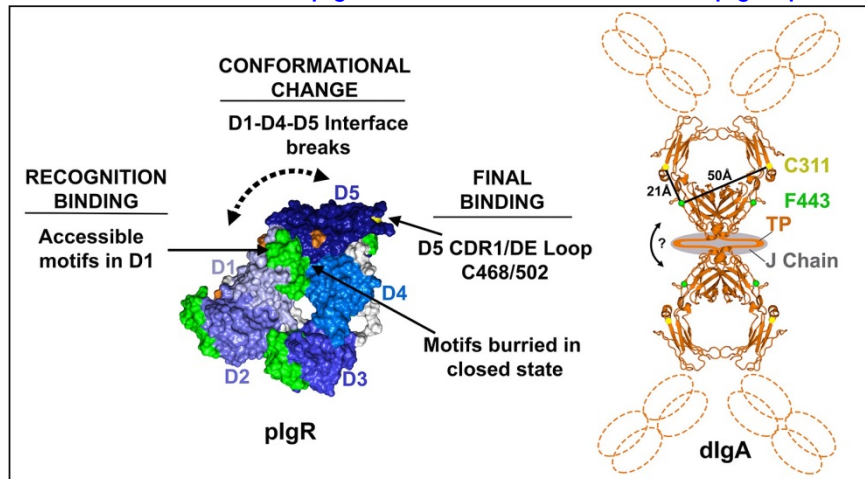
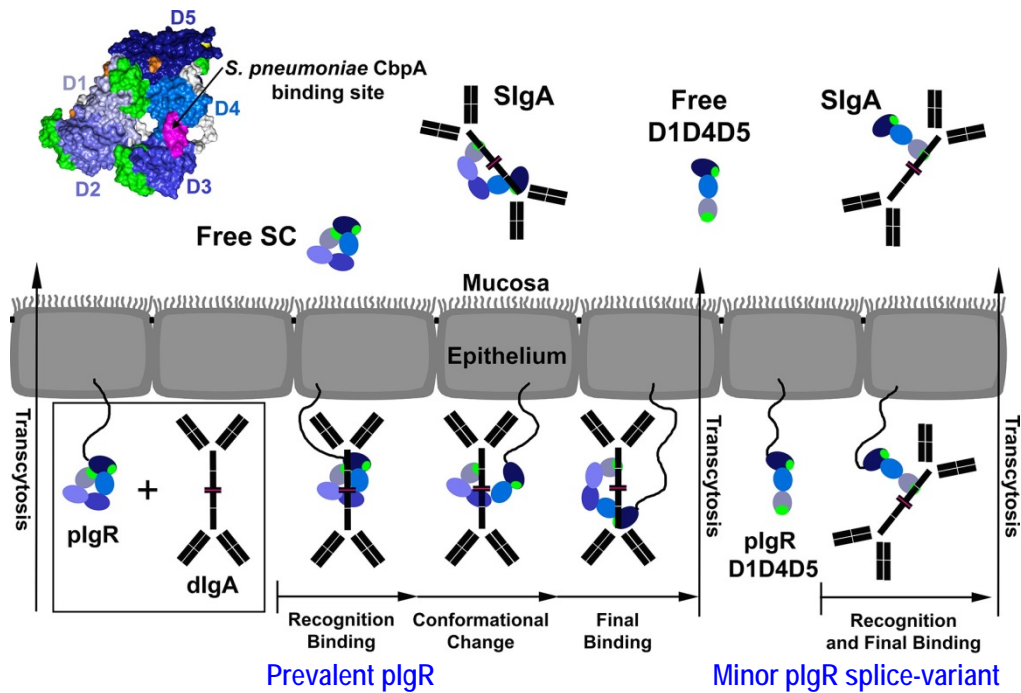
Not found in Small mammals
Higher monkeys

T-shape
Less-Sensitive to proteases

Secretory : planar (fab fragments
aligned with Fc portion)

No binding to Peyer's Patch M
cells. Warning! Assay used mouse
M cells.
Rochereau N *et al.* PLoS Biol 2013

Poly Ig Receptor (pIgR), Secretory component (SC) and plgs



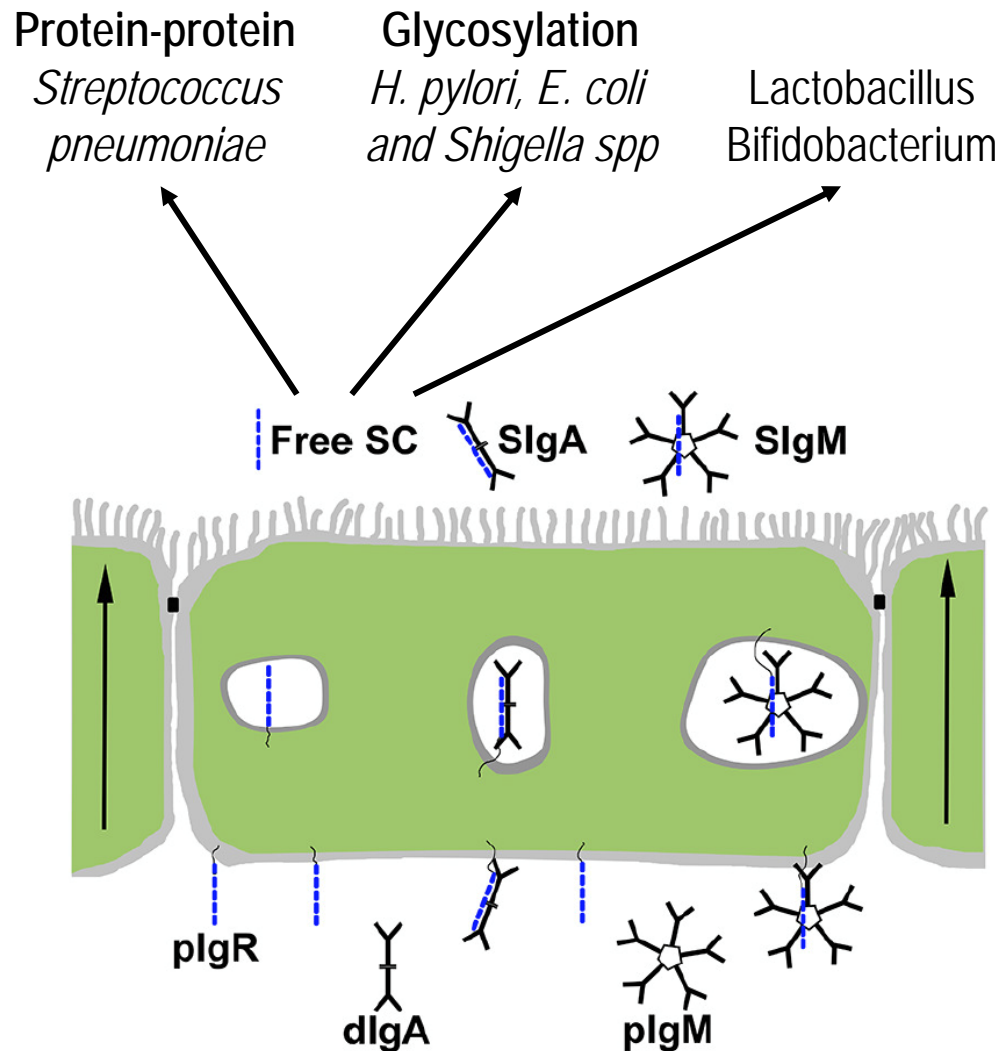
Human pIgR:

- Glycosylated type I membrane protein
- Five tandem immunoglobulin-like (Ig-like) domains (620AA)
- Transmembrane domain (23AA)
- Intracellular domain (103AA)

pIgR in evolution

- Oldest identifiable Fc receptor,
- Teleost (bony) fish (2 Ig-like domains)
- Birds, amphibians and reptiles (4 Ig-like domains)
- Mammals (5 Ig-like domains (D1-D5)) plus a splice variant (D1D4D5)

Poly Ig Receptor (pIgR), Secretory component (SC) and plgs



Secretory component

- <50% of pIgR transcytose in empty state.
- Free SC may bind bacteria and toxins.
 - *Streptococcus pneumoniae* (protein-protein)
 - *H. pylori, E. coli and Shigella spp* (glycosylation)
 - Lactobacillus and Bifidobacterium (unknown binding mode)
- Free SC may bind mucus (glycosylation dependent).
- Free SC binds pIgM 10x stronger than dIgA (maybe not identical to pIgR affinity)

pIgR – dimeric-IgA (dIgA)

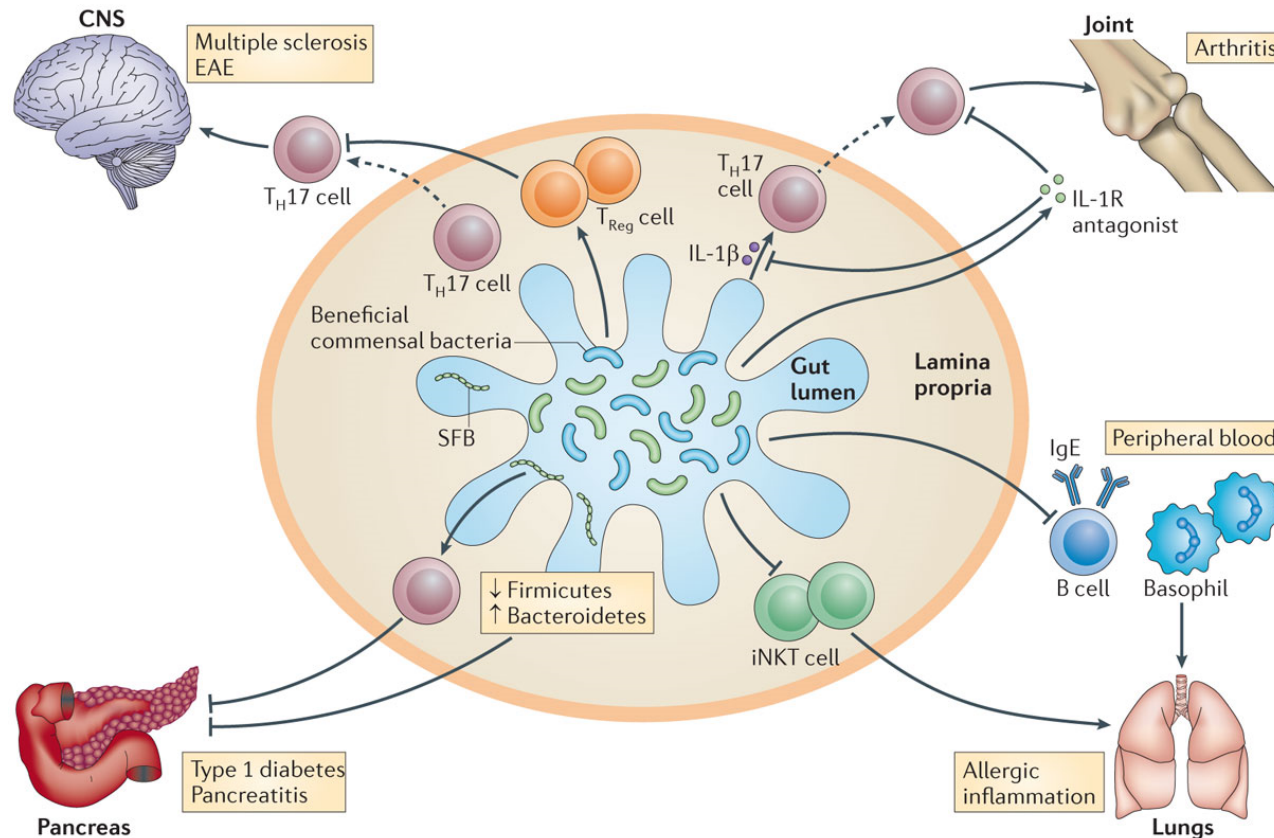
- Bound by pIgR of all tetrapods (J-chain is equally conserved).
- Bound by D1 followed by D5 (D5 is finally covalently attached)

pIgR – pentameric-IgM

- Bound by D1 but not D5.
- Not covalently linked

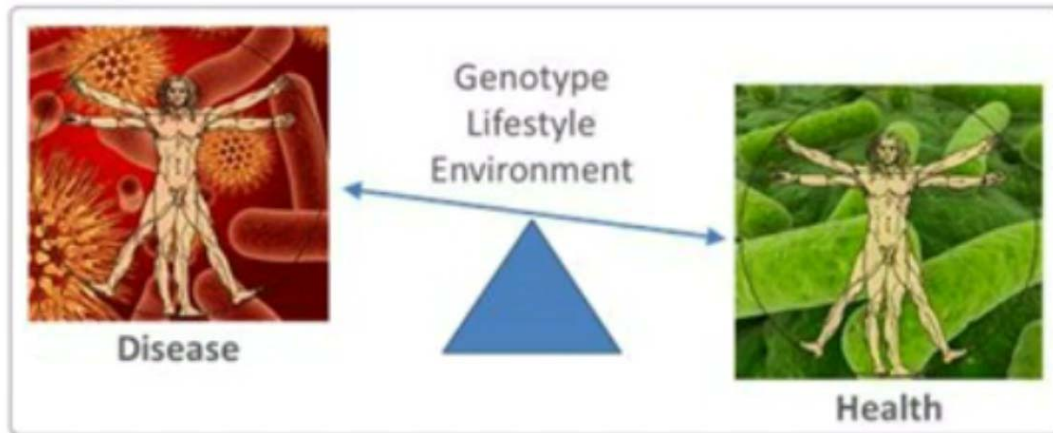
Gut microbiota and autoimmunity

Autoimmunity associated with gut microbiota

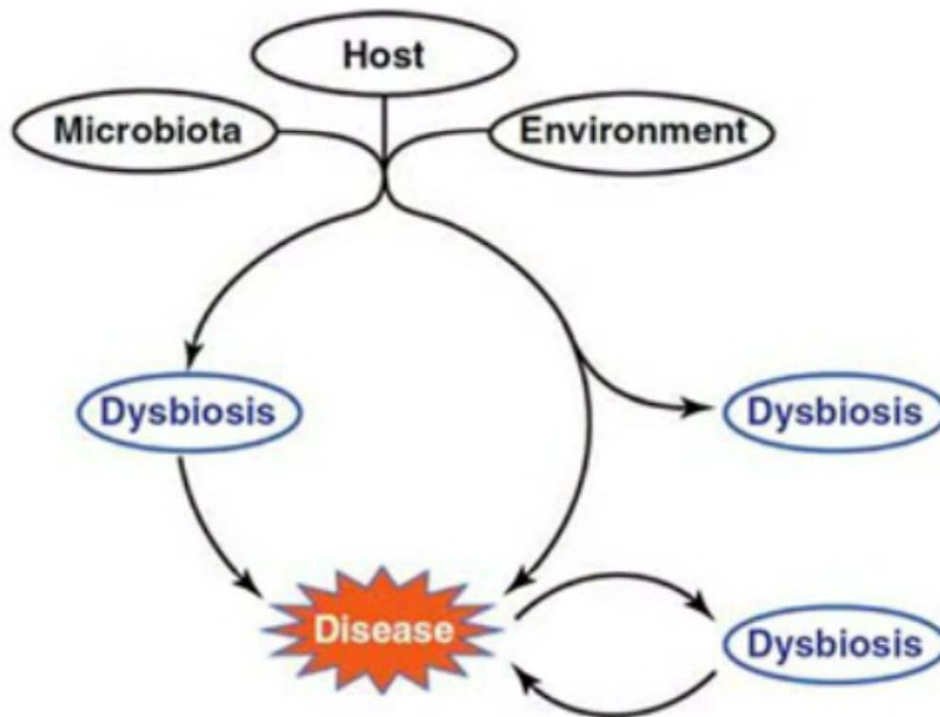


- IBD ([Arumugam et al. Nature 2011](#), [Juste et al. Gut 2014](#), [Palm et al. Cell 2014](#))
- Type-1 Diabetes ([Qin et al. Nature 2012](#), [Markle et al. Science 2013](#))
- Arthritis ([Scher et al. Nat Rev Rheumatol 2011](#), [Scher et al. eLife 2013](#))
- Allergy ([Russell et al. EMBO Rep 2012](#))
- **EAE / Multiple sclerosis** ([Berer et al. Nature 2011](#), [Miyake et al. PLoS One 2015](#))

Dysbiosis - cause or consequence of disease?



- Genetic or environmental factors may lead to dysbiosis
- Dysbiosis may lead to disease
- Genetic or environmental factors may lead to disease irrespective of dysbiosis.
- Disease may lead to dysbiosis



Koch's postulate and why it doesn't always apply

Koch's Postulates

(Robert Koch and Friedrich Loeffler in 1884)

Evidence required to establish etiologic relationship between microorganism and disease:

- Microorganism must be observed in every case of the disease

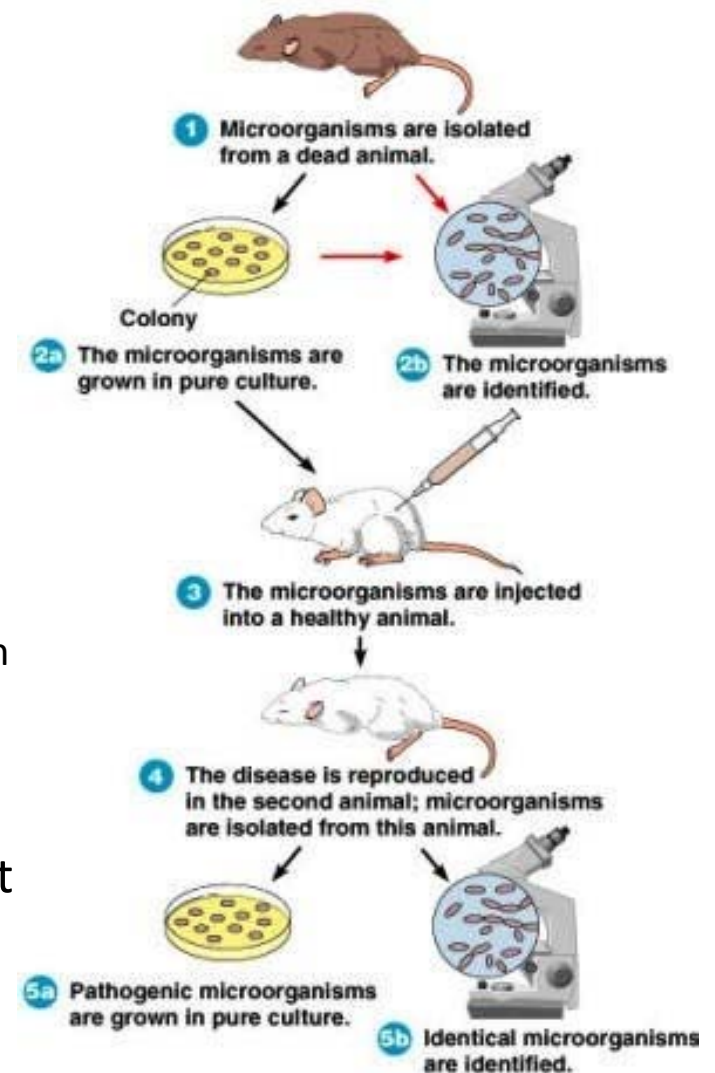
Criticism: Healthy carriers exist (Cholera, Typhoid, but also viruses like Zoster and HIV)

- It must be isolated and grown in pure culture

Criticism: Not all microbes can be cultivated and viruses only in presence of their host. Effective vaccines eradicating e.g. polio is considered a good proof of the causality of polio virus.

- The pure culture, when inoculated in animals, must reproduce the disease

- Microorganism must be recovered from the diseased animal.



Bradford Hill criteria - epidemiological alternative to Koch

Bradford Hill Criteria

(Epidemiologist Sir Austin Bradford Hill in 1965)

- **Strength (effect size):** A small association does not mean that there is not a causal effect, though the larger the association, the more likely that it is causal.
- **Consistency (reproducibility):** Consistent findings observed by different persons in different places with different samples strengthens the likelihood of an effect.
- **Specificity:** Causation is likely if there is a very specific population at a specific site and disease with no other likely explanation. The more specific an association between a factor and an effect is, the bigger the probability of a causal relationship.
- **Temporality:** The effect has to occur after the cause (and if there is an expected delay between the cause and expected effect, then the effect must occur after that delay).
- **Biological gradient:** Greater exposure should generally lead to greater incidence of the effect. However, in some cases, the mere presence of the factor can trigger the effect. In other cases, an inverse proportion is observed: greater exposure leads to lower incidence.
- **Plausibility:** A plausible mechanism between cause and effect is helpful (but Hill noted that knowledge of the mechanism is limited by current knowledge).
- **Coherence:** Coherence between epidemiological and laboratory findings increases the likelihood of an effect. However, Hill noted that "... lack of such [laboratory] evidence cannot nullify the epidemiological effect on associations".
- **Experiment:** "Occasionally it is possible to appeal to experimental evidence".
- **Analogy:** The effect of similar factors may be considered.

Dysbiosis - cause or consequence - and so what?

“Le malade imaginaire”

Molière, 1673

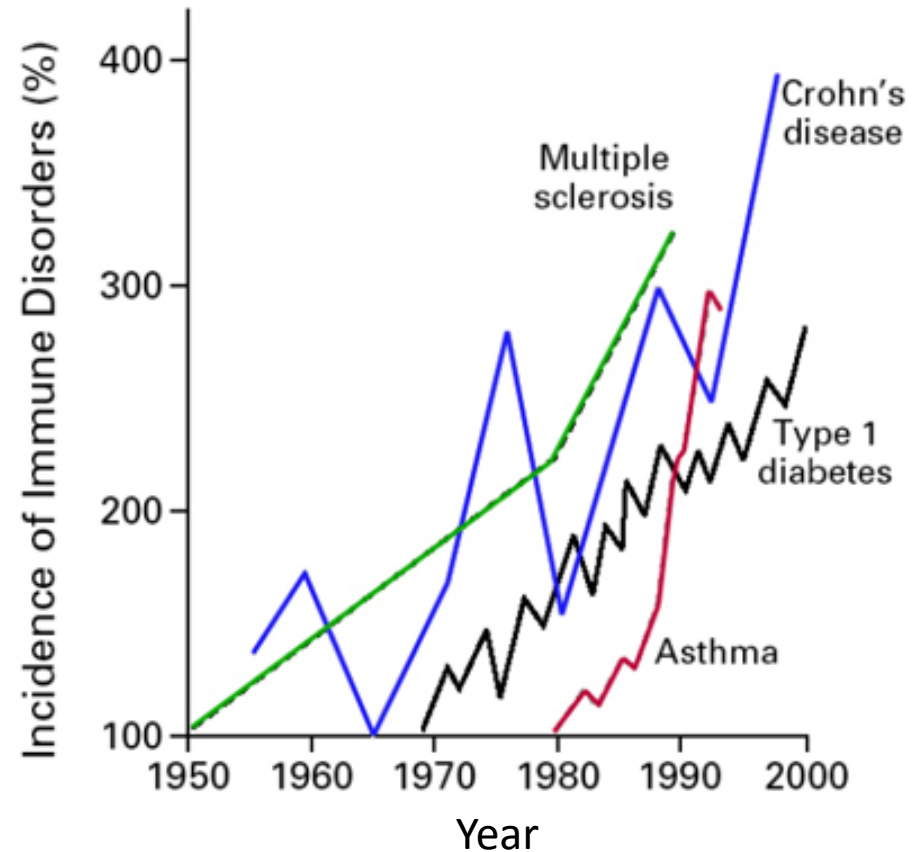
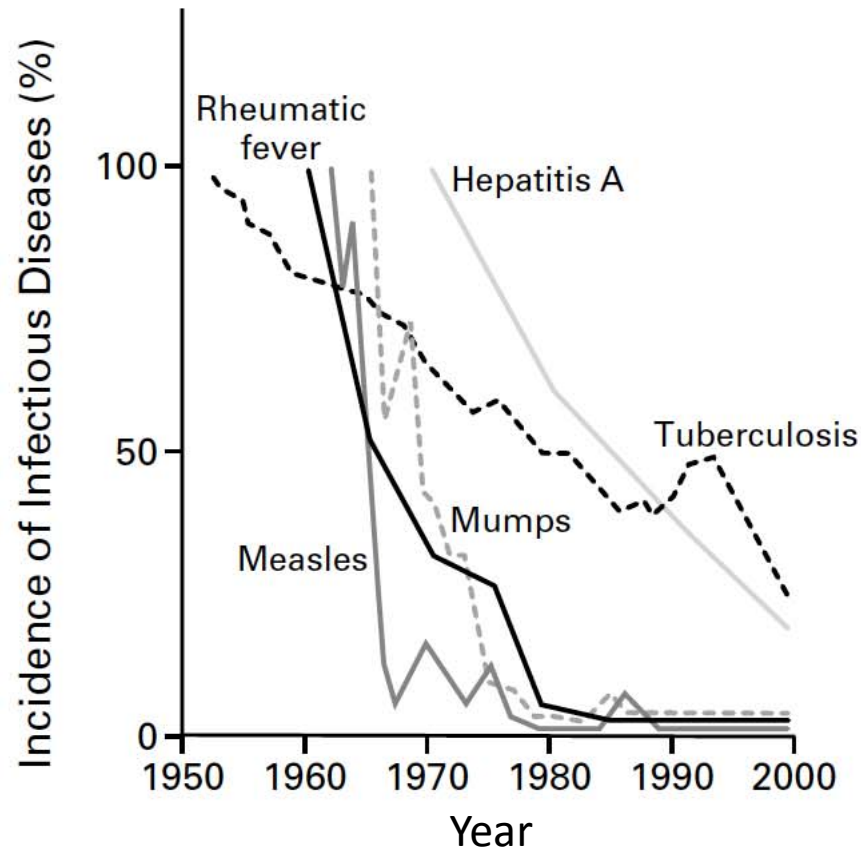


- Prescription of “lavement” for:
 - Digestion, intestinal secretion and bad mood.
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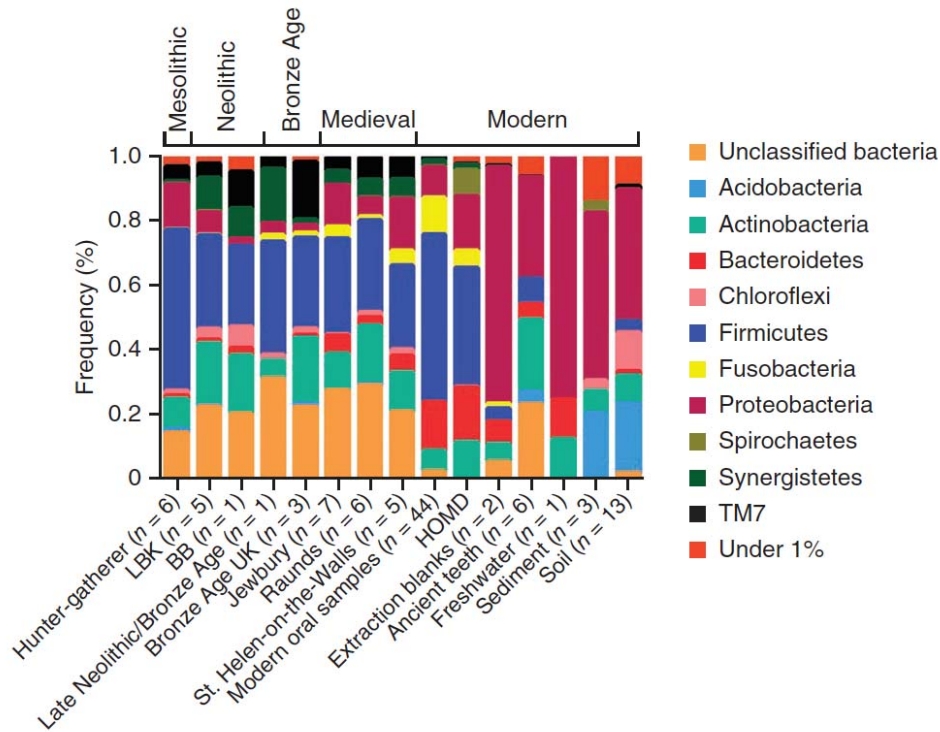
- The bi-directional interactions between gut microbiota, metabolic and endocrine functions of the organism suggest that impacting one will impact the other.
- If the gut microbiota is not the cause:
 - Treatment targeting the microbiota will not be curative,
 - but may temporarily cure symptoms.
 - Many treatments actively used are non-curative (e.g. HIV therapy)
- If the gut microbiota is the cause:
 - Treatment targeting the microbiota is curative (*Clostridium difficile* infections),

Hygiene theory and autoimmunity



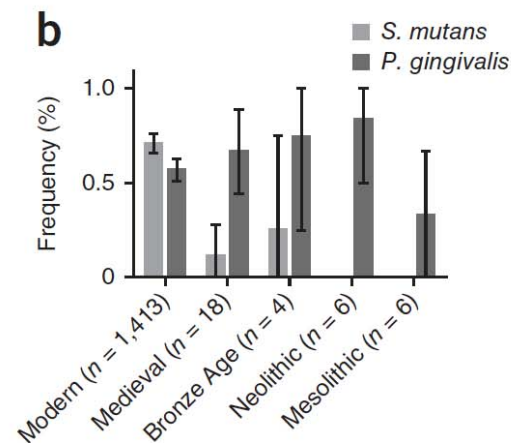
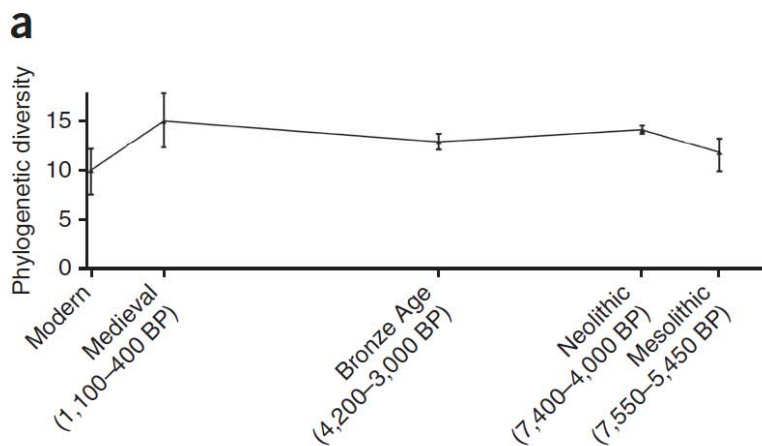
Disappearance of prototypic infectious diseases inversely correlate with occurrence of autoimmune disease.

Oral microbiota shifts through history of mankind

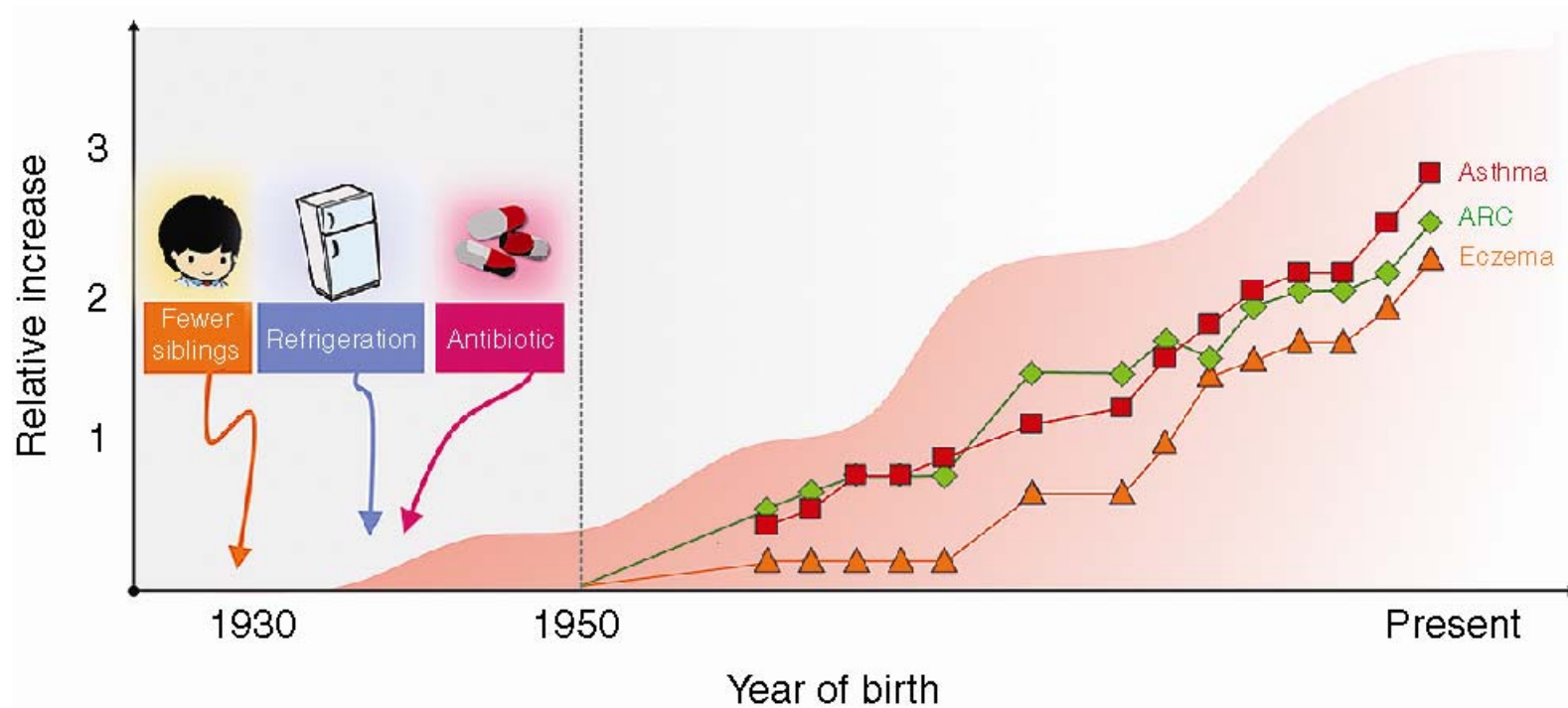


Two of the greatest dietary shifts in human evolution involved :

- Adoption of carbohydrate-rich Neolithic (farming) diets (beginning ~10,000 years before the present)
 - *P. gingivalis* augments.
- Industrially processed flour and sugar (in ~1850)
 - *Streptococcus mutans* augments (associated with carries).
- Reduced oral microbiota diversity in the modern human.



Microbiota shifts in modern history



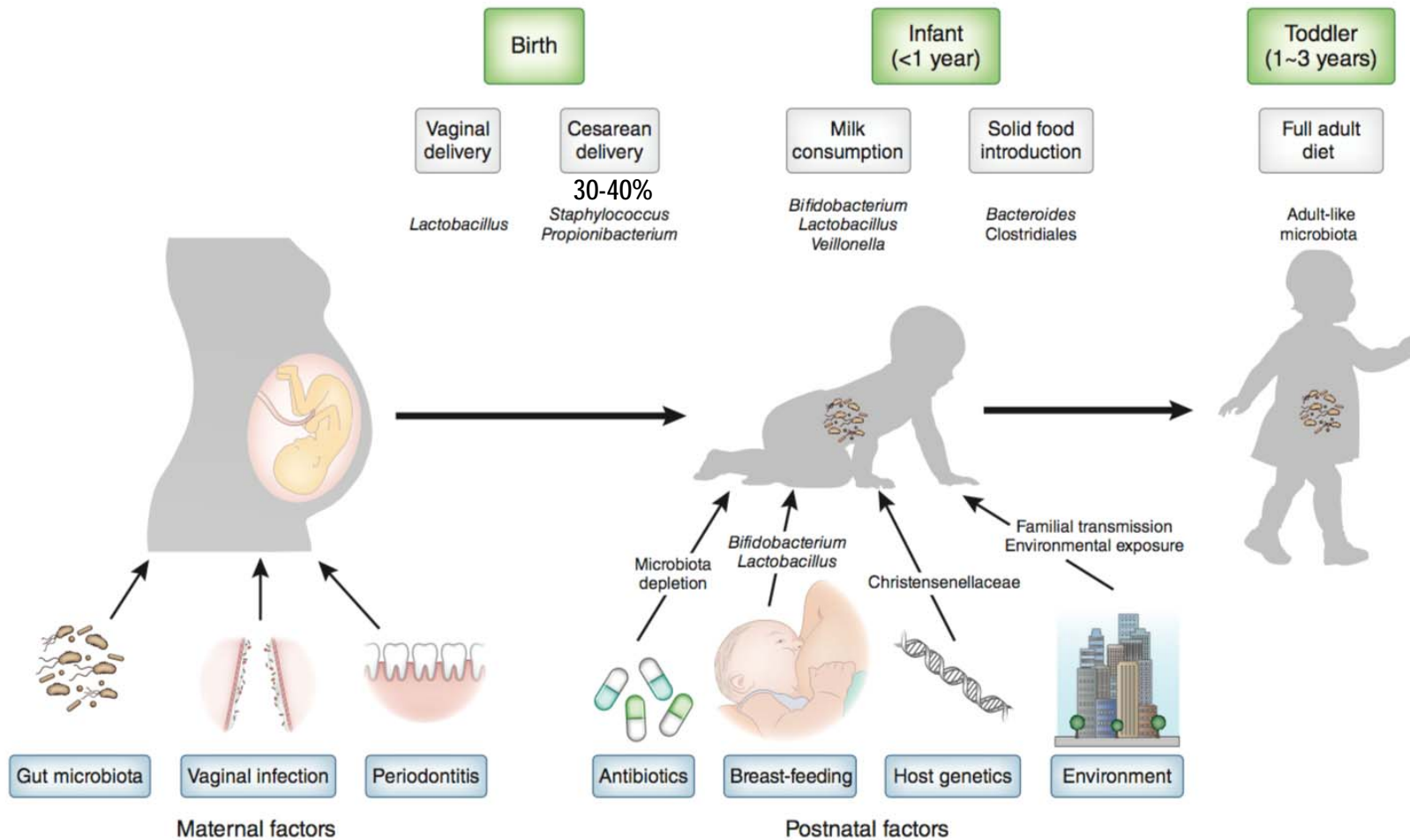
- Recent lifestyle alterations:
 - Fewer siblings
 - Refrigerators (cleaner food)
 - Antibiotics

Increase in allergy

Lifestyle changes affecting Gut Microbiota

- Massive increase in prevalence of allergic diseases in Westernized countries (>20% over 10 year period)
- Allergic disease is attributed to both genetic predisposition and environmental factors
- Genetic drift over such a short period of time cannot explain increased incidence of disease
- Westernized life-style has introduced several environmental risk factors that disturb the homeostatic balance of gut microbiota
- Excessive antibiotic use, especially during early life (or even during pregnancy)
- Shift towards more formula-fed babies
- Shift towards greater numbers of babies born by Caesarean section
- Western diet

Early-life factors affecting infant gut microbiota

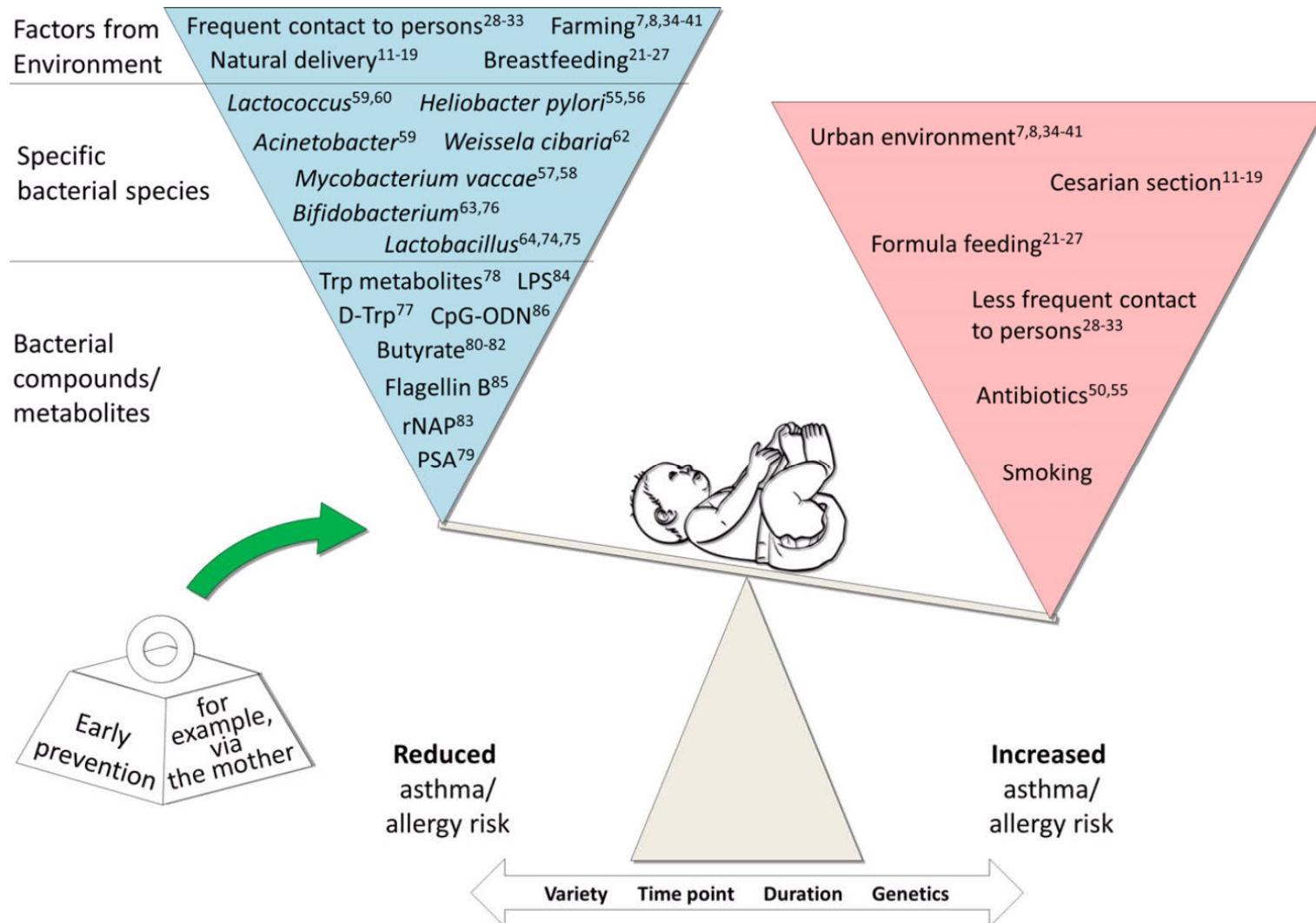


- Both maternal and environmental factors influence early-life gut microbiota colonization.

C-section and allergy



C-section and allergy



C-section and allergy

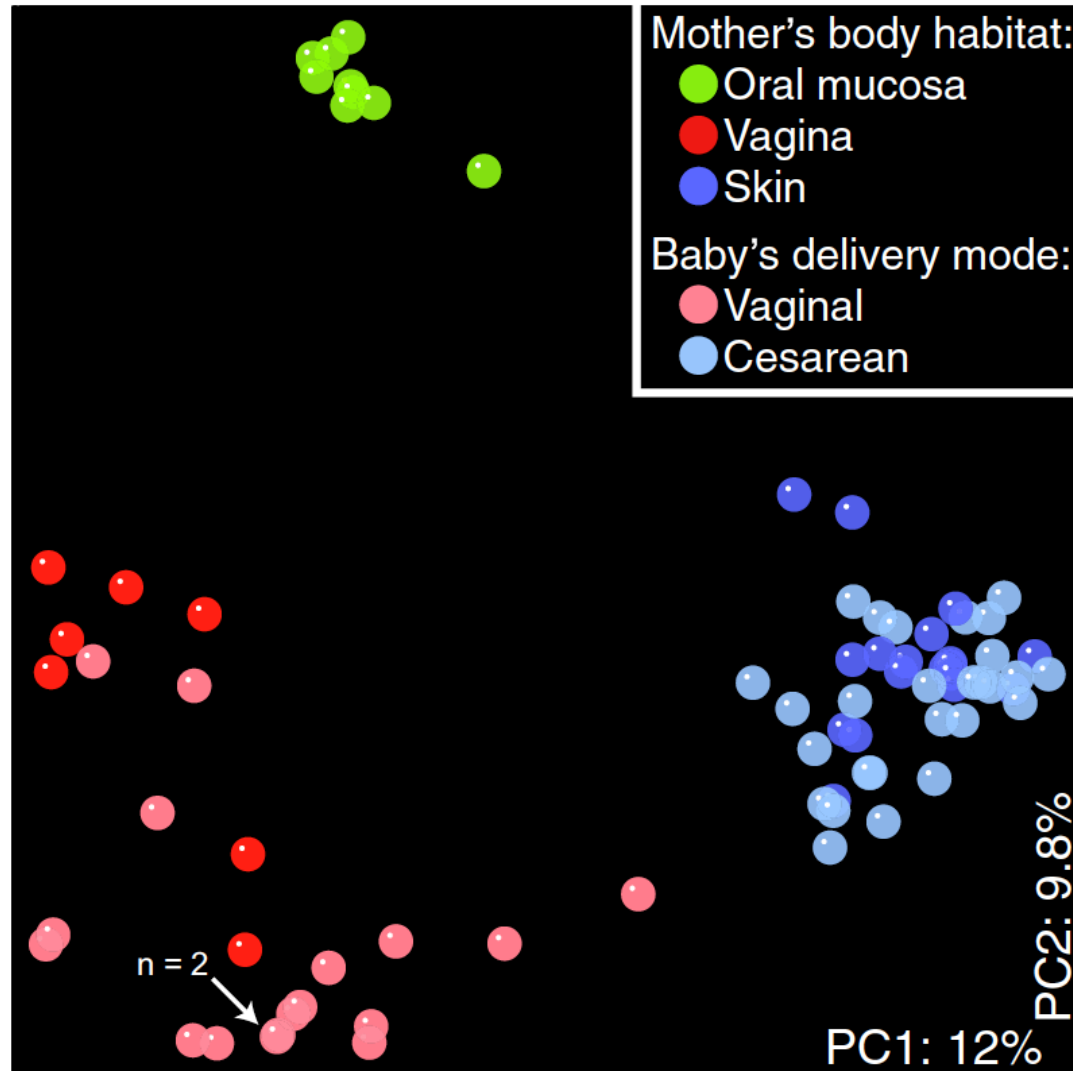
- **Obesity:** Caesarean born babies are at double the risk of becoming obese.
- **Allergy:** Associated with elective/planned C-section (odds ratio=1.49 [1.13-1.97]). Not significant for emergency C-sections (n>60.000).
 - An intact membrane (more frequent in elective C-section) is associated with allergy. Breaking the membrane may result in the first bacterial exposure.

Sevelsted et al. J Pediat 2016; Rusconi et al. Am J Epid 2017
- **Asthma:** Elective C-section (OR = 1.58 [1.17-2.13], n=1400).
 - Exclusive Breastfeeding for 6 months (OR = 1.39 [0.92-2.10]).
 - Non-exclusive breastfeeding or bottle feeding (OR = 1.91 [1.22-2.99]).

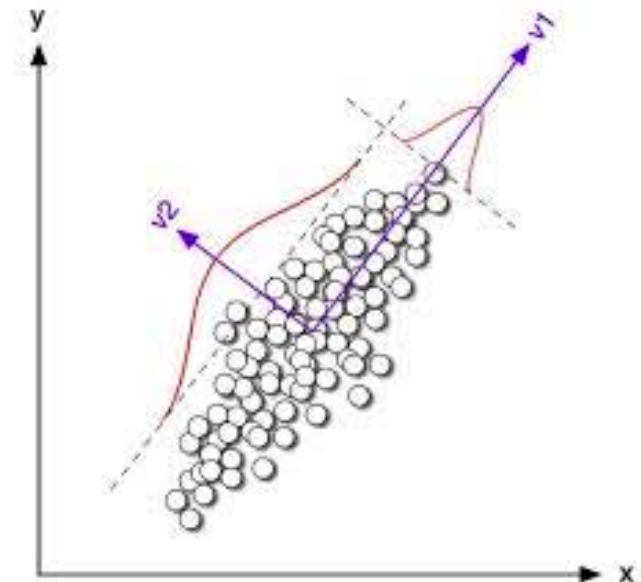
Chu et al. PLoS One 2017
- **Gut colonization** at 1 week:
 - C-section: *Citrobacter freundii*, *Clostridium* species, *Enterobacter cloacae*, *Enterococcus faecalis*, *Klebsiella oxytoca*, *Klebsiella pneumoniae*, and *Staphylococcus aureus*
 - Vaginal: *Escherichia coli*
 - Differences disappear before Age 1.
 - Initial airway microbiota was unaffected by birth method.

Stockholm et al. JACI 2016

Delivery mode shape early life gut microbiote colonization



Principal component analysis
Define axis representing maximal data variance!!



Vaginal Microbial Transfer (VMT) rescue microbiota post c-section.

nature
medicine

Partial restoration of the microbiota of cesarean-born infants via vaginal microbial transfer

Maria G Dominguez-Bello^{1,2}, Kassandra M De Jesus-Laboy², Nan Shen³, Laura M Cox¹, Amnon Amir⁴, Antonio Gonzalez⁴, Nicholas A Bokulich¹, Se Jin Song^{4,5}, Marina Hoashi^{1,6}, Juana I Rivera-Vinas⁷, Keimari Mendez⁷, Rob Knight^{4,8} & Jose C Clemente^{3,9}

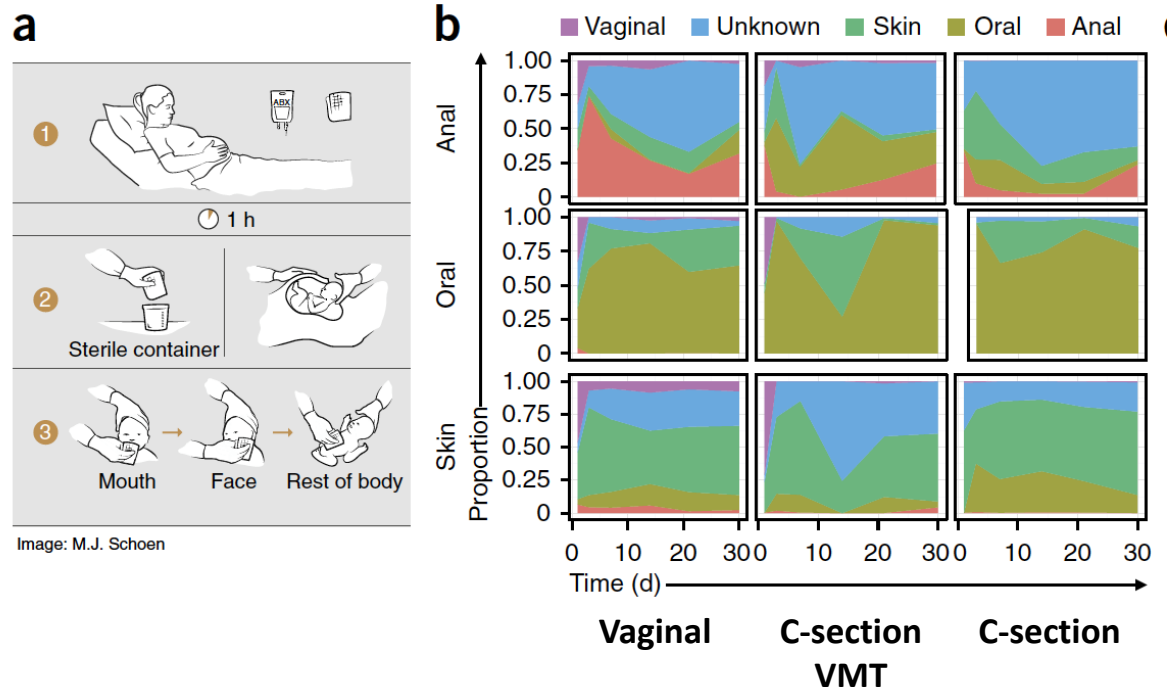


Image: M.J. Schoen

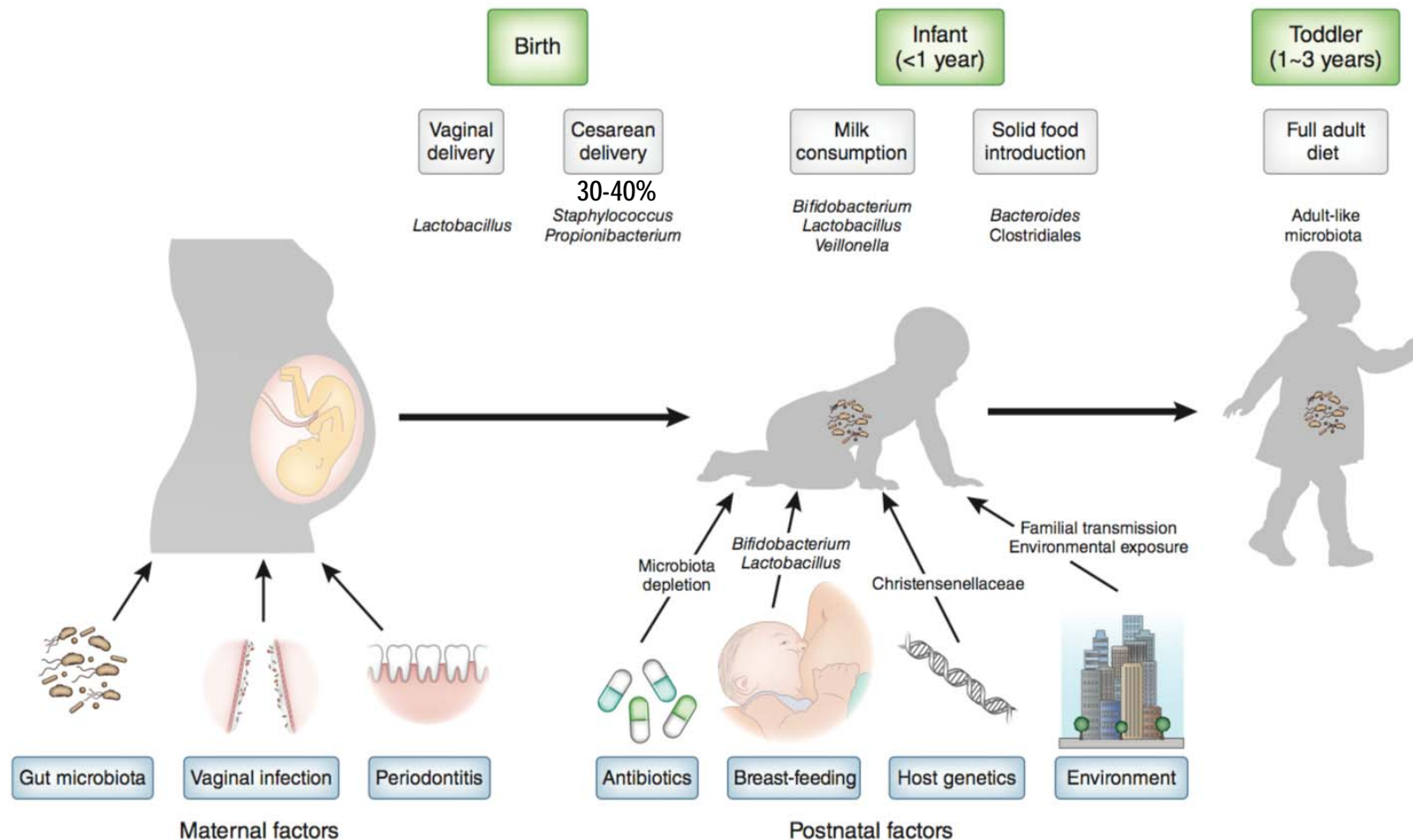
- Vaginal Microbial Transfer (VMT) partly rescue gut microbiota composition of children born by C-section.
- Would primarily be maternal vaginal microbiota (mother/baby paired).
- However, in the case of mothers treated with antibiotics (such as HIV infected mothers) allogenic microbiota may be of interest – even for children born vaginally.

Early-life factors affecting infant gut microbiota

| | Children age | | | | | | | | Mother | | | |
|-------------------------------|--------------|------|----------|------|----------|------|----------|-------------|----------|------|-------------|------|
| | 6 days | | 3 weeks | | 2 months | | 6 months | | Before | | After birth | |
| | <i>n</i> | SI | <i>n</i> | SI | <i>n</i> | SI | <i>n</i> | SI | <i>n</i> | SI | <i>n</i> | SI |
| Lifestyle | | | | | | | | | | | | |
| Antroposopic | 45 | 1.45 | 44 | 1.43 | 49 | 1.31 | 47 | 1.35 | 50 | 2.59 | 51 | 2.58 |
| Not | 65 | 1.47 | 57 | 1.41 | 64 | 1.33 | 62 | 1.64 | 66 | 2.59 | 65 | 2.60 |
| Living on a farm | | | | | | | | | | | | |
| No | 91 | 1.45 | 84 | 1.41 | 94 | 1.34 | 91 | 1.63 | 98 | 2.60 | 99 | 2.59 |
| Yes | 19 | 1.49 | 17 | 1.53 | 19 | 1.30 | 18 | 1.25 | 18 | 2.48 | 17 | 2.58 |
| Mother vegetarian | | | | | | | | | | | | |
| No | 99 | 1.47 | 90 | 1.41 | 100 | 1.33 | 97 | 1.63 | 103 | 2.60 | 104 | 2.58 |
| Yes | 11 | 1.45 | 11 | 1.50 | 13 | 1.31 | 12 | 1.10 | 13 | 2.58 | 12 | 2.73 |
| Antibiotics pregnancy | | | | | | | | | | | | |
| No | 97 | 1.39 | 87 | 1.43 | 98 | 1.30 | 95 | 1.60 | 100 | 2.60 | 99 | 2.59 |
| Yes | 13 | 1.81 | 14 | 1.26 | 15 | 1.34 | 14 | 1.53 | 16 | 2.57 | 16 | 2.53 |
| Birthplace | | | | | | | | | | | | |
| Hospital* | 73 | 1.38 | 68 | 1.40 | 75 | 1.32 | 70 | 1.52 | – | – | – | – |
| Home | 21 | 1.35 | 19 | 1.39 | 20 | 1.40 | 21 | 1.36 | – | – | – | – |
| Birthmode | | | | | | | | | | | | |
| Vaginal | 94 | 1.37 | 87 | 1.39 | 95 | 1.33 | 91 | 1.52 | – | – | – | – |
| Caesarean | 16 | 1.59 | 14 | 1.62 | 18 | 1.24 | 18 | 1.70 | – | – | – | – |
| Sex | | | | | | | | | | | | |
| Boy | 51 | 1.48 | 44 | 1.41 | 53 | 1.30 | 53 | 1.49 | – | – | – | – |
| Girl | 59 | 1.40 | 57 | 1.41 | 60 | 1.36 | 56 | 1.62 | – | – | – | – |
| Milk formula 1st week | | | | | | | | | | | | |
| Yes | 27 | 1.33 | 25 | 1.35 | 28 | 1.29 | 29 | 1.78 | – | – | – | – |
| No | 89 | 1.45 | 80 | 1.39 | 89 | 1.30 | 87 | 1.49 | – | – | – | – |
| Breastfeeding 2 months | | | | | | | | | | | | |
| Exclusive | – | – | – | – | 94 | 1.30 | 88 | 1.39 | – | – | – | – |
| Partly | – | – | – | – | 14 | 1.58 | 15 | 1.84 | – | – | – | – |
| Not | – | – | – | – | 5 | 1.66 | 6 | 2.14 | – | – | – | – |
| Breastfeeding 6 months | | | | | | | | | | | | |
| Exclusive | – | – | – | – | – | – | 25 | 1.32 | – | – | – | – |
| Partly | – | – | – | – | – | – | 67 | 1.60 | – | – | – | – |
| Not | – | – | – | – | – | – | 17 | 1.94 | – | – | – | – |
| All samples | 110 | 1.49 | 101 | 1.44 | 113 | 1.35 | 109 | 1.61 | 116 | 2.46 | 116 | 2.48 |

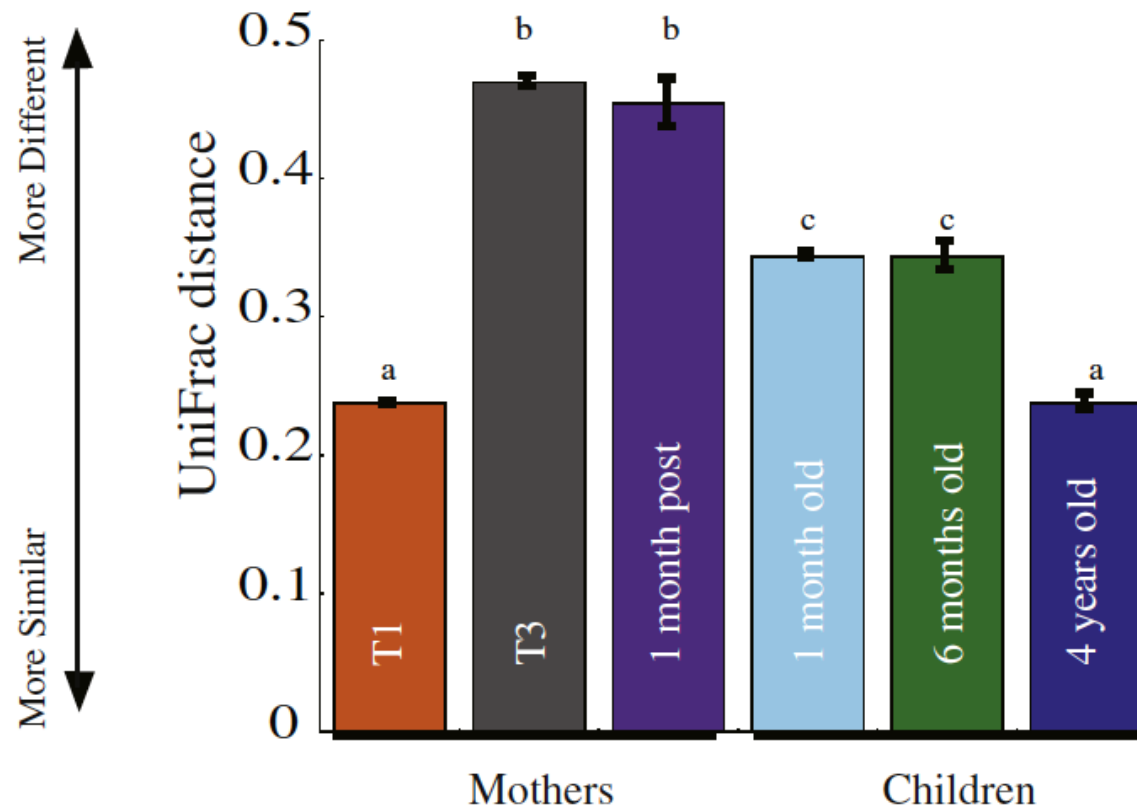
- **Birth route:** C-section associated with increased diversity in early-life
- **Breastfeeding:** Microbiota diversity reduced at 6 months.
- **Mother diet:** Vegetarians => reduced diversity at 6 months

Early-life factors affect infant gut microbiota temporarily



- But at 1-3 years of age the child acquire an adult like gut microbiota, which is identical for all “healthy” children, independent of birth route and breastfeeding.
- If gut microbiota is unaltered, but disease risk elevated – what can be the cause.
- Gut microbiota immunity?

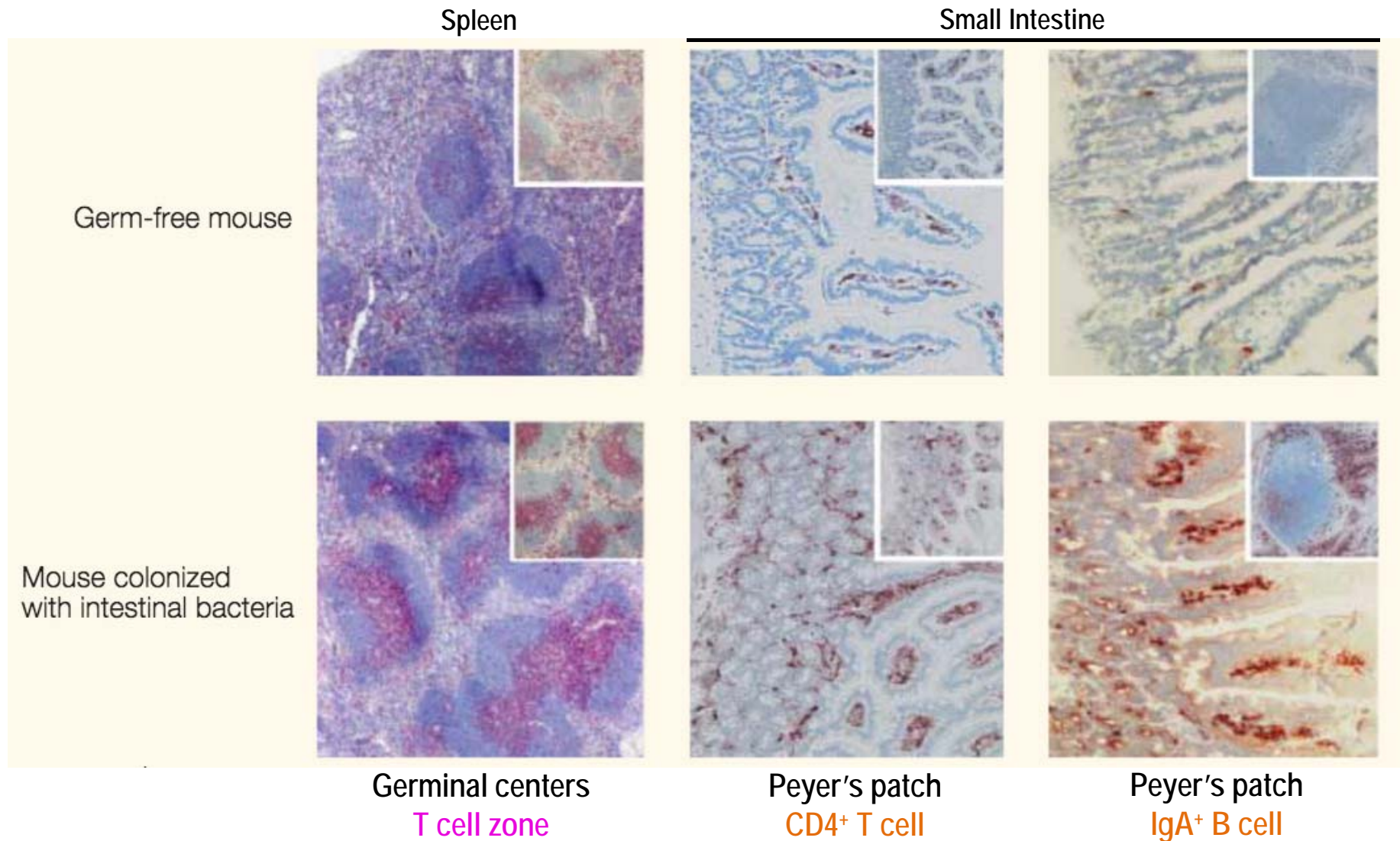
Microbiota alterations during pregnancy and early life



- Maternal microbiota changes between 1st and 3rd trimester and remains altered until at least 1 month post birth.
- Infant microbiota is different from initial T1 maternal gut microbiota.
- Children approaches the maternal microbiota at 4 years of age.

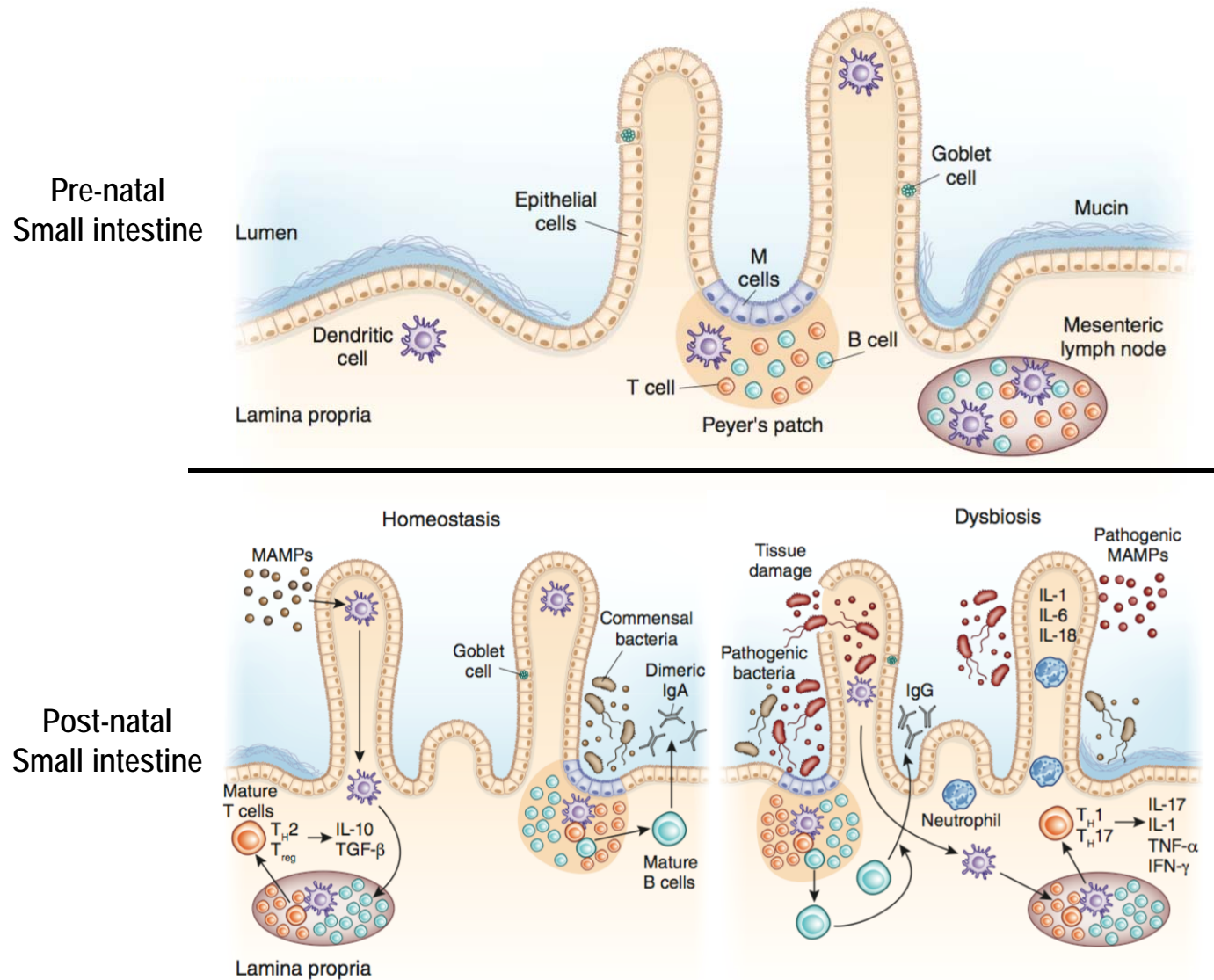
What happens to immunity?

Lymphoid structures and immune cells in GF mice



- Spleen with few germinal centres and poorly formed T cell (pink) and B cell zones.
- Germ-free mice display hypoplastic Peyer's patches, with reduced T cell numbers and IgA-expressing B cells.

Early-life factors affecting infant gut immunity and health



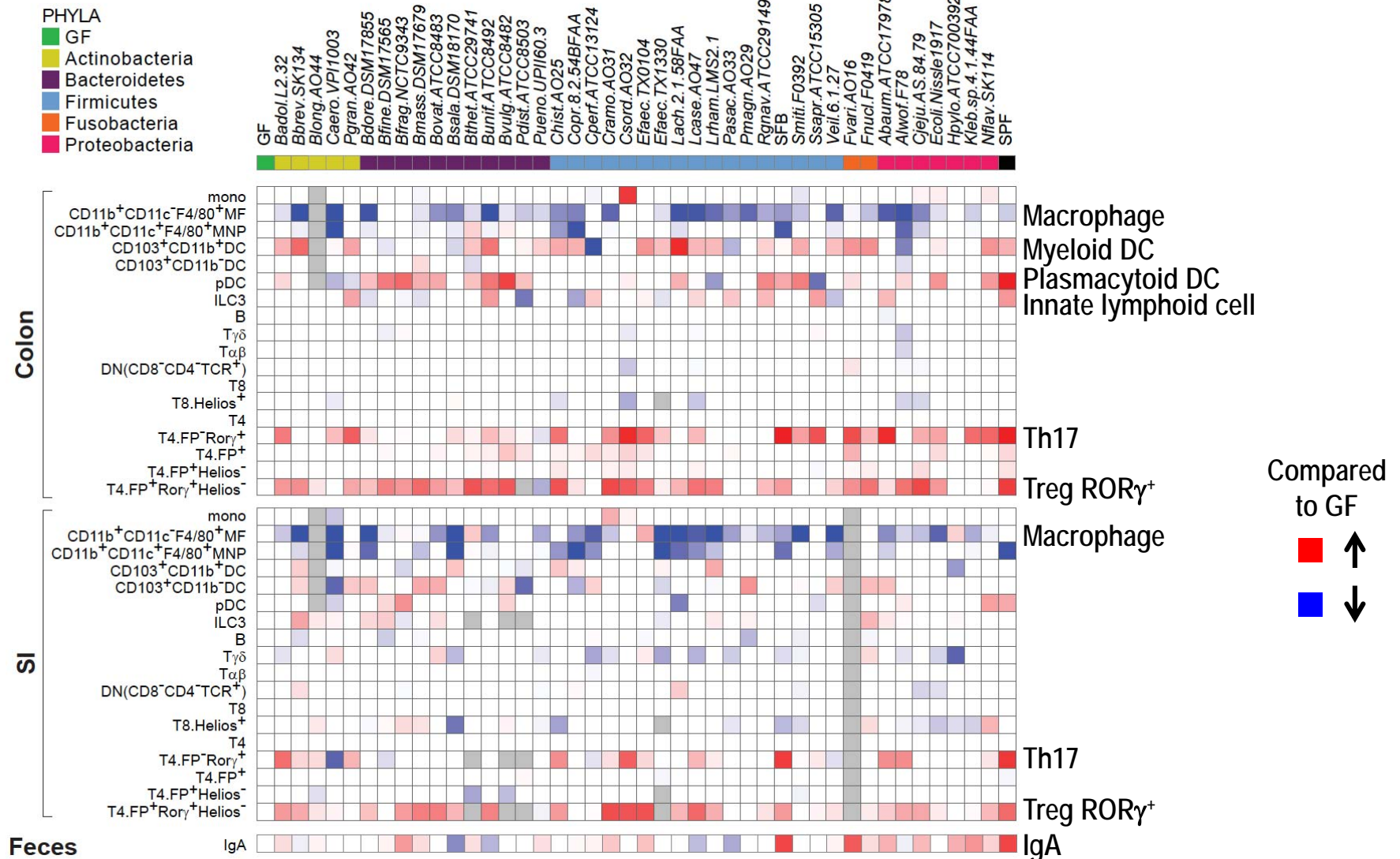
- Of note, Pre-natal Peyer's patches are still fairly unstructured with weak definition of B and T cell zones of the lymphoid follicle.

Microbes stimulate T cell immunity



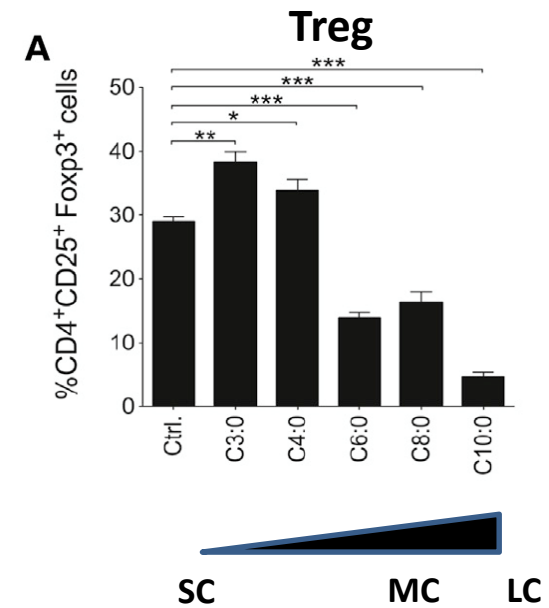
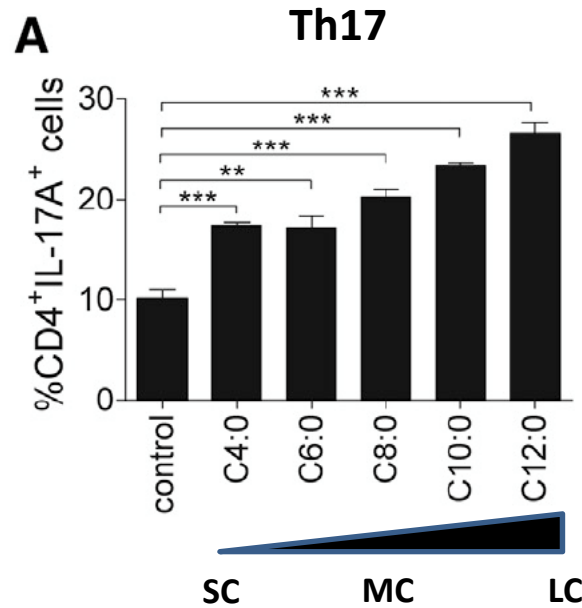
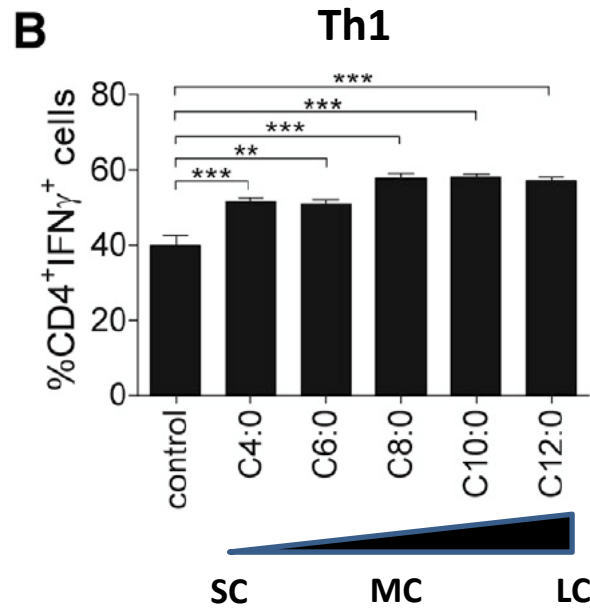
- Monocolonization of GF mice with 53 bacterial strains.
- Extensive immune phenotyping

Microbes stimulate T cell immunity



- APCs shifts from macrophages to DCs (more professional antigen presentation)
- Commensals increase gut residing Th17 and Treg cells.
- Phylum independent strain specific effects (e.g. free IgA in feces).

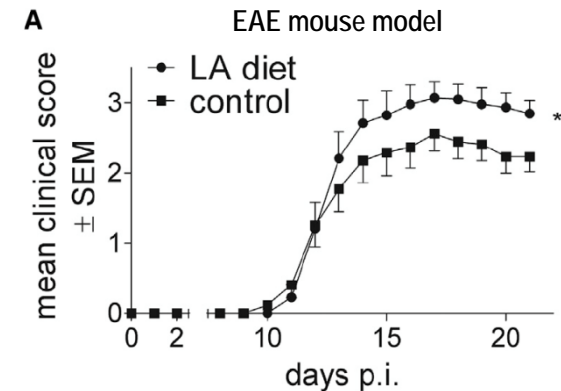
Remark! Metabolites also affect host immunity



Short-chain (SC) versus Long-chain (LC) fatty acids.

| | Treg | Th1/Th17 | EAE |
|-------------------------|------|----------|-----|
| Propionate (PA, C3) | ↑ | ↓ | ↓ |
| Lauric Acid (LA, C12) | ↓ | ↑ | ↑ |
| Palmitic Acid (PA, C16) | ↓ | ↑ ↑ | ↑ ↑ |

Fatty acids control T cell differentiation, not growth rate.



Immuno-Microbiota Interactions

TABLE 12-12 · LEVELS OF IMMUNOGLOBULINS IN SERA OF NORMAL SUBJECTS BY AGE*

| Age | IgG | | IgM | | IgA | | Total Immunoglobulin | |
|----------|-------------------------|------------------|---------|------------------|----------|------------------|----------------------|------------------|
| | mg/dl | % of Adult Level | mg/dl | % of Adult Level | mg/dl | % of Adult Level | mg/dl | % of Adult Level |
| Newborn | 1031 ± 200 [†] | 89 ± 17 | 11 ± 5 | 1.1 ± 5 | 2 ± 3 | 1 ± 2 | 1044 ± 201 | 67 ± 13 |
| 1-3 mo | 430 ± 119 | 37 ± 10 | 30 ± 11 | 30 ± 11 | 21 ± 13 | 11 ± 7 | 481 ± 127 | 31 ± 9 |
| 4-6 mo | 427 ± 186 | 37 ± 16 | 43 ± 17 | 43 ± 17 | 28 ± 18 | 14 ± 9 | 498 ± 204 | 32 ± 13 |
| 7-12 mo | 661 ± 219 | 58 ± 19 | 54 ± 23 | 55 ± 23 | 37 ± 18 | 19 ± 9 | 752 ± 242 | 48 ± 15 |
| 13-24 mo | 762 ± 209 | 66 ± 18 | 58 ± 23 | 59 ± 23 | 50 ± 24 | 25 ± 12 | 870 ± 258 | 56 ± 16 |
| 25-36 mo | 892 ± 183 | 77 ± 16 | 61 ± 19 | 62 ± 19 | 71 ± 37 | 36 ± 19 | 1024 ± 205 | 65 ± 14 |
| 3-5 yr | 929 ± 228 | 80 ± 20 | 56 ± 18 | 57 ± 18 | 93 ± 27 | 47 ± 14 | 1078 ± 245 | 69 ± 17 |
| 6-8 yr | 923 ± 256 | 20 ± 22 | 65 ± 25 | 66 ± 25 | 124 ± 45 | 62 ± 23 | 1112 ± 293 | 71 ± 20 |
| 9-11 yr | 1124 ± 235 | 97 ± 20 | 79 ± 33 | 80 ± 33 | 131 ± 60 | 66 ± 30 | 1334 ± 254 | 85 ± 17 |
| 12-16 yr | 946 ± 124 | 82 ± 11 | 59 ± 20 | 60 ± 20 | 148 ± 63 | 74 ± 32 | 1153 ± 169 | 74 ± 12 |
| Adults | 1158 ± 305 | 100 ± 26 | 99 ± 27 | 100 ± 27 | 200 ± 61 | 100 ± 31 | 1457 ± 353 | 100 ± 24 |

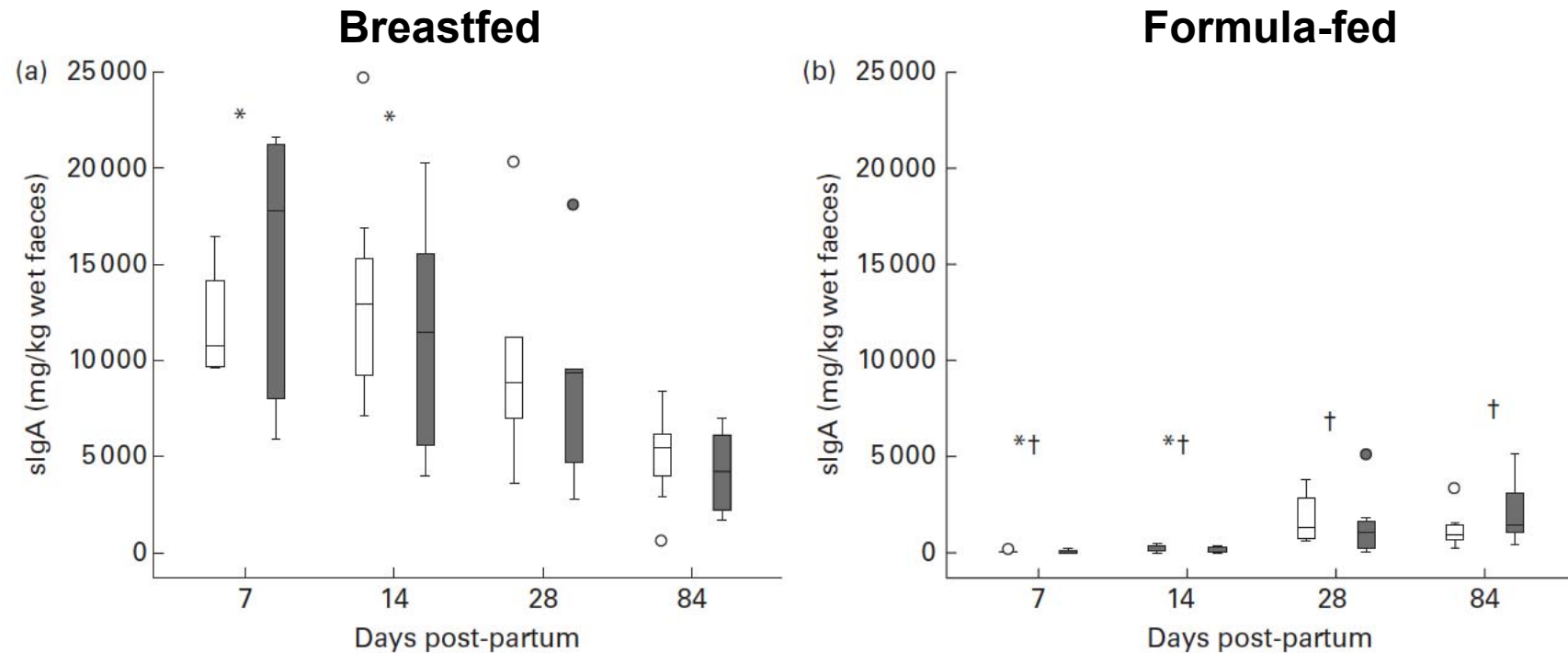
*The values were divided from measurements made in 296 healthy children and 30 adults. Levels were determined by the radial diffusion technique using specific rabbit antisera to human immunoglobulins.

[†]One standard deviation.

From Stiehm ER, Fudenberg HH. Serum levels of immune globulins in health and disease. A survey. *Pediatrics* 37:715, 1966.

- We are born with maternal IgG antibodies circulating our blood stream.
- IgM and IgA **serum** antibodies are virtually absent at birth and slowly increases during childhood.
- Intestinal antibodies are provided through breastfeeding.

IgA- Microbiota interactions

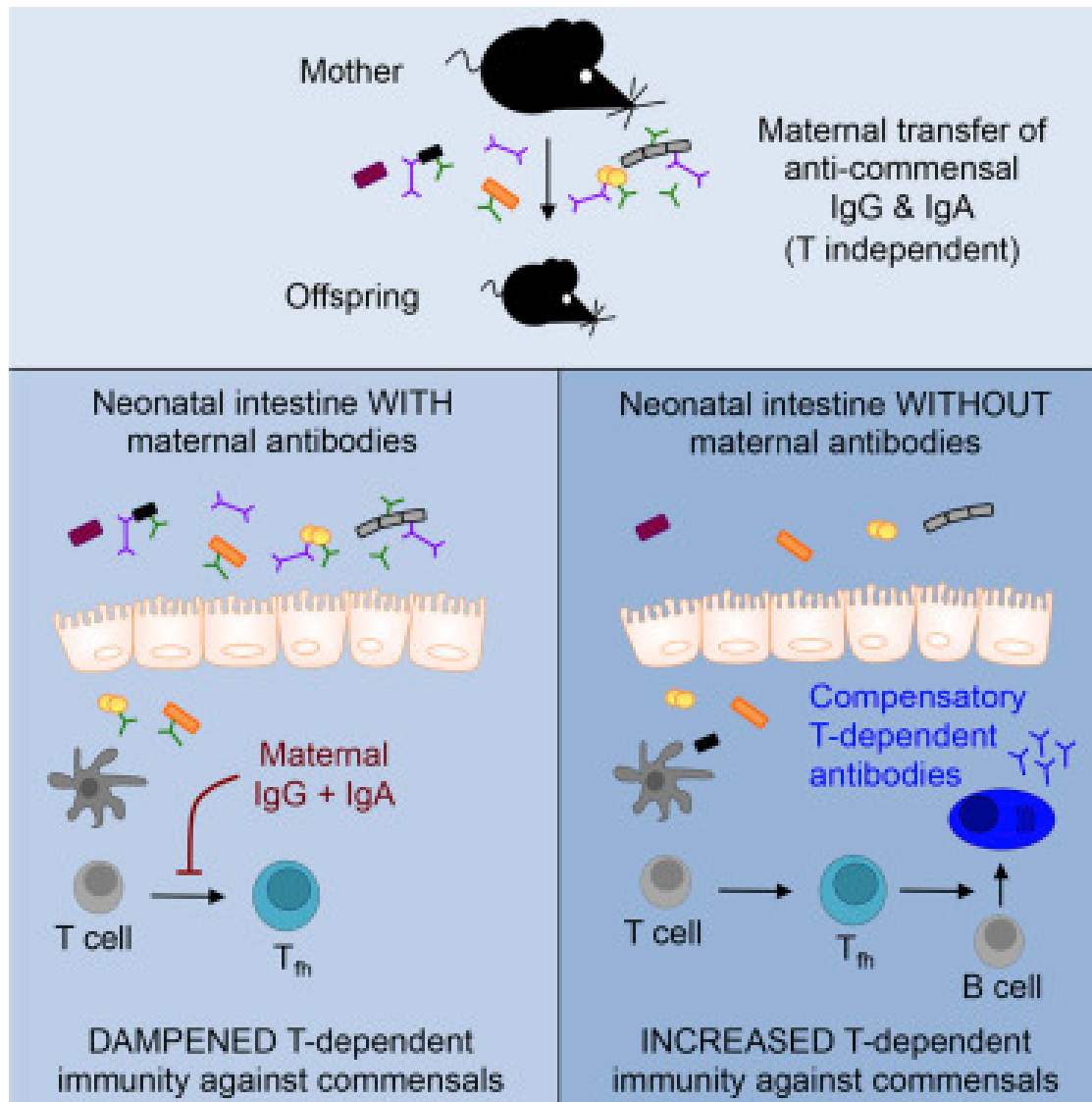


sIgA is high in breastfed infant feces and decreases over-time, likely depicting the fact that the milk becomes less rich in sIgA (colostrum (1st milk) versus breastmilk).

Formula-fed infants have initially no sIgA in their feces, but at one month post partum small amounts of autologous sIgA appear in their feces.

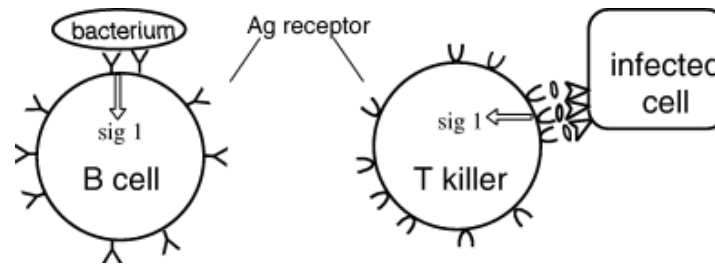
Women on Salmon diet (black bars) versus normal diet (white bars). Diet has no impact.

Maternal antibodies dampen offspring immunity

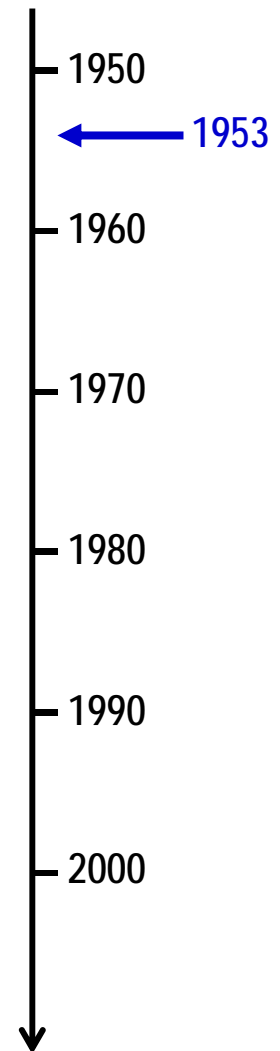


- T-independent (no class-switch) IgG specific for mucosal bacteria.
- Gut microbes elicit anti-commensal IgG antibodies via TLR signalling on B cells
- Maternal transmission of IgG coordinates with IgA to limit mucosal T cell responses
- Absence of maternal antibodies triggers a compensatory T-dependent immune response in the offspring.

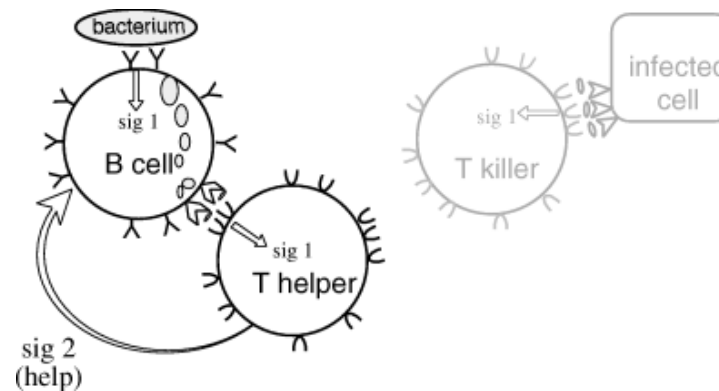
Self/non-self versus Danger model in a historical perspective



- Burnet and Lederberg propose the antigen receptor (BCR and TCR)
- Antigen stimulation (signal 1) induces immunity including class switch and somatic hyper mutation.

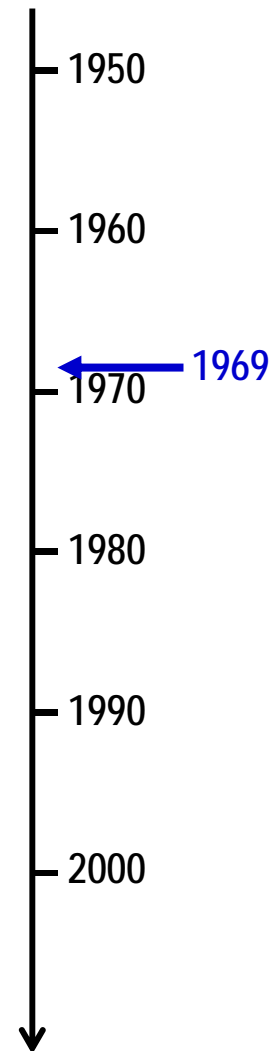


Self/non-self versus Danger model in a historical perspective

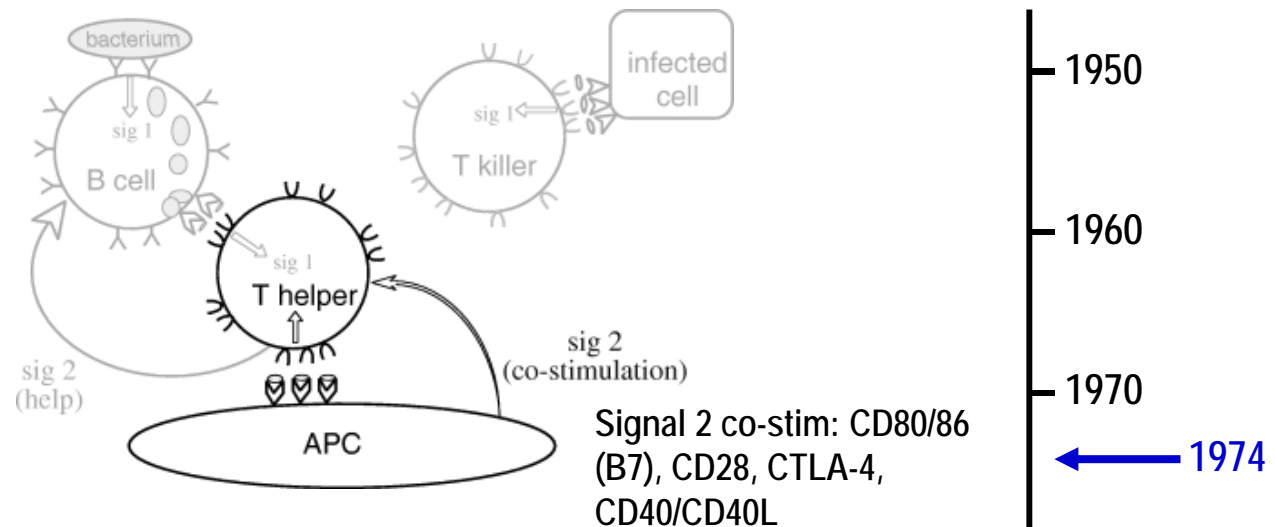


Signal 2: e.g. CD40/CD40L and cytokines

- **Problem:** BCR hypermutation may lead to autoreactive BCRs.
- **Solution:** Cohn add another cell: The T helper cell (only formally proven much later).
- B cells internalize pathogen and present antigens to interact with specific Th cells, which validate that target is non-self.
- Signal 1 alone leads to clonal deletion (both self and non-self reactivity leads to signal 1).
- Signal 1 + 2 lead to activation (non-self rescued by T cells, with TCR which does not hyper mutate).

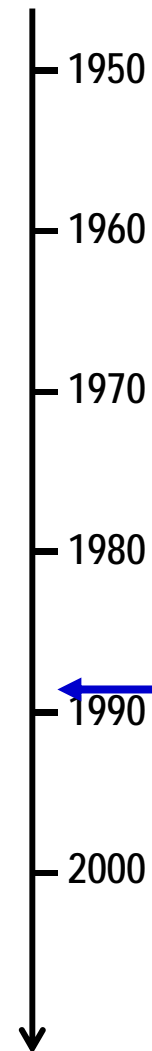
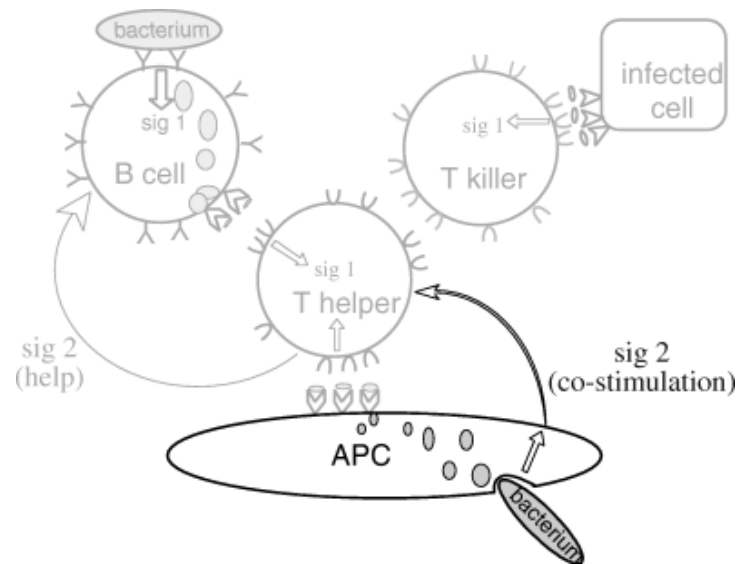


Self/non-self versus Danger model in a historical perspective



- **Problem:** We need a BCR independent manner to capture and present antigens to prime naïve T cells to become T helper cells.
- **Solution:** Lafferty and Cunningham propose that Th cells are primed and activated through APC antigen presentation and co-stimulation.
- Th cells are not constitutive active – feedback regulation upon antigen removal.
- Heavily criticized because APCs do not explain how the immune system distinguish between self and non/self (which BCR dependent antigen selection provided).

Self/non-self versus Danger model in a historical perspective



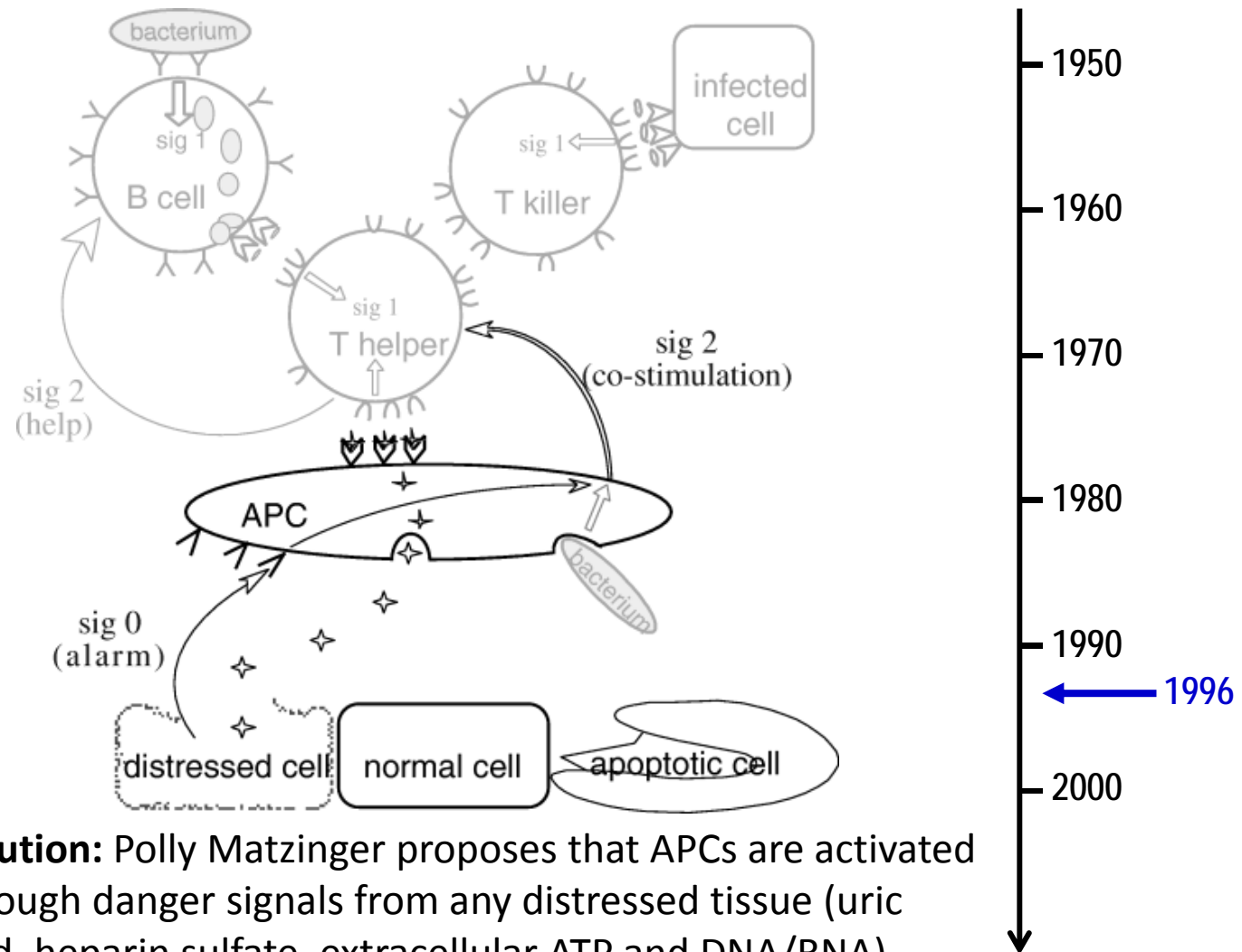
- **Problem:** Contrary to B cells, APCs do not differentiate between self and non-self.
- **Solution:** Charlie Janeway proposes that APCs internalize and present non-self selectively through **pattern recognition receptors (PRR)**, which bind elements from foreign organisms, such as bacteria.
- This model inherently imply that APCs are not constitutively active, but require external stimuli through the PRR signalling pathway.
- Propose explanation why vaccines need an adjuvant.

Self/non-self versus **Danger** model in a historical perspective

Problem

How to explain:

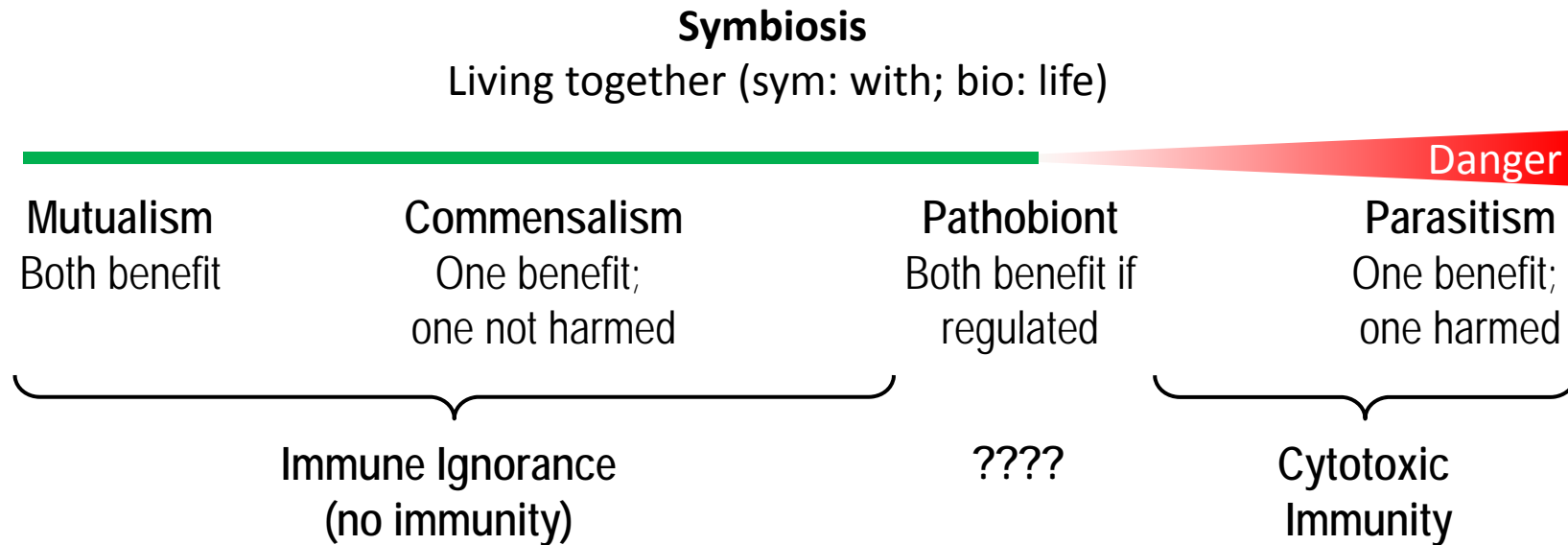
- Autoimmunity
- Non-reject of tumour with tumour antigen
- DNA therapy versus DNA vaccine
- Why mothers don't reject the fetus.
- Why temporal gene-expression changes doesn't evoke immunity (e.g. breast milk).
- Why can we host tons of microbes?



- **Solution:** Polly Matzinger proposes that APCs are activated through danger signals from any distressed tissue (uric acid, heparin sulfate, extracellular ATP and DNA/RNA).
- Advocate that we are a friendly host as long as our visitors are friendly too. Don't push the button first policy.

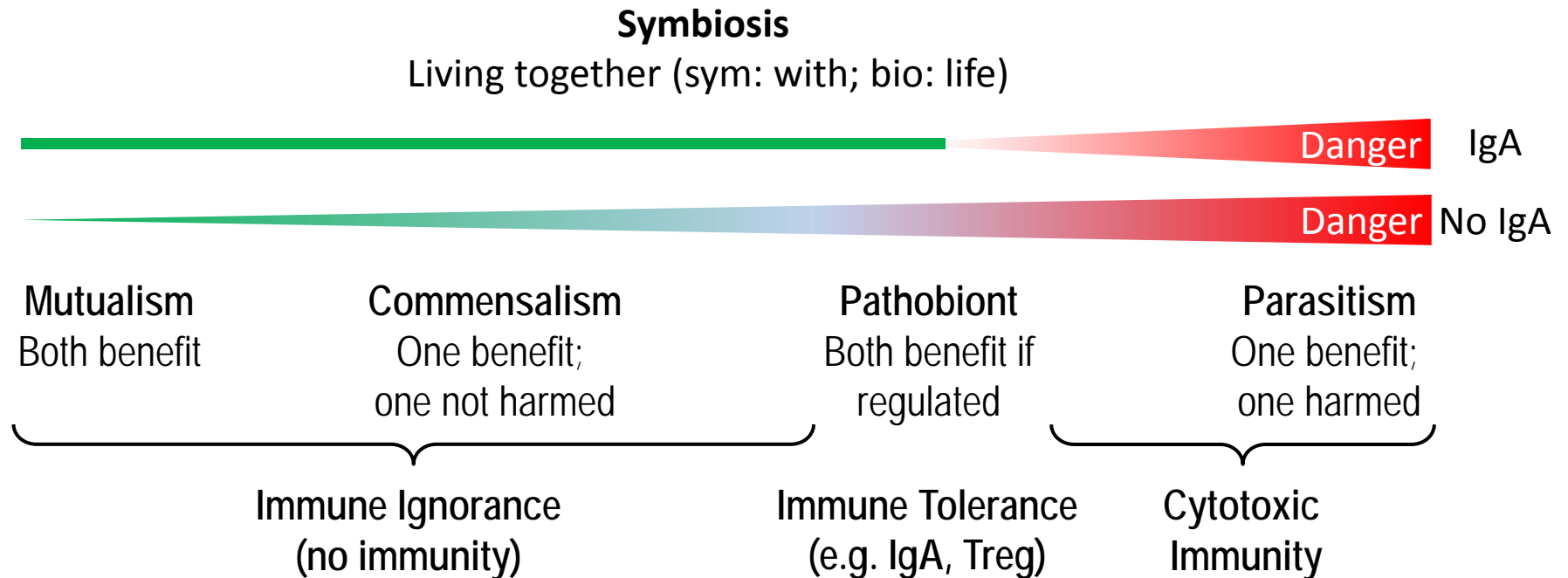
Danger model and gut microbiota symbiosis

- The danger model would suggest that we do not respond to non-harmful bacteria colonizing our gut (Mutualism and Commensalism allowed).



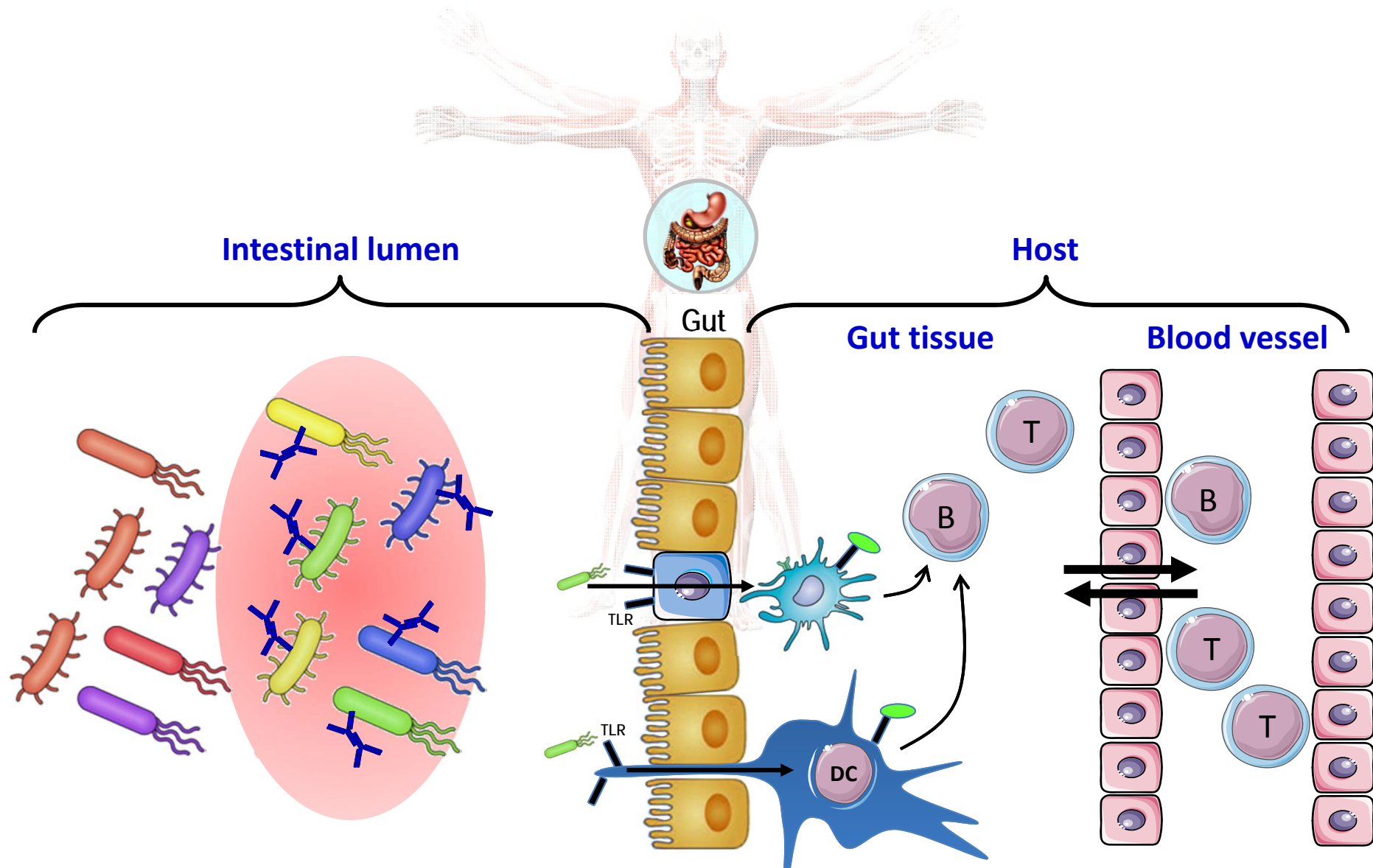
Danger model and IgA responses

- The danger model would suggest that we do not respond to non-harmful bacteria colonizing our gut (Mutualism and Commensalism allowed).

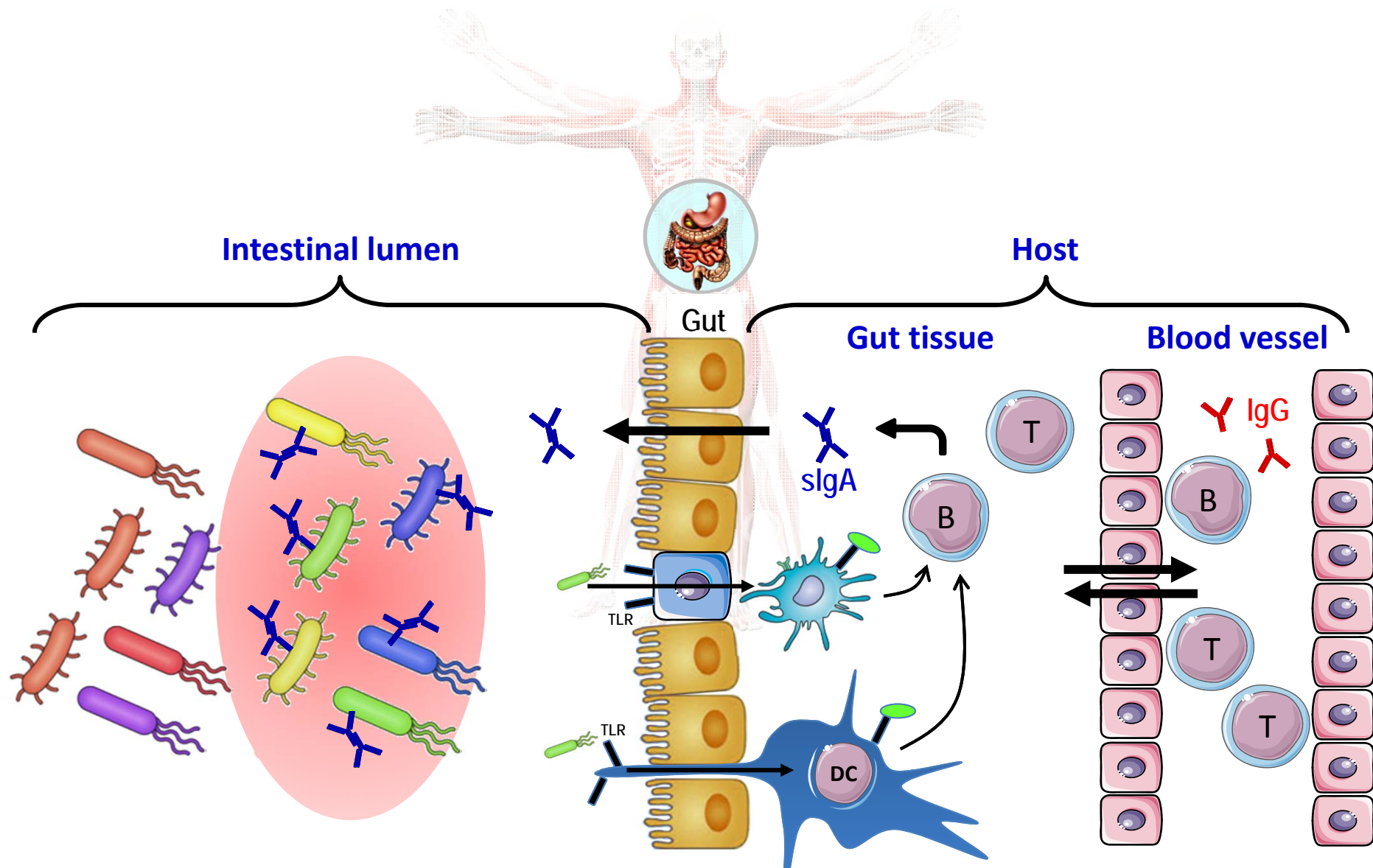


- In reality microbes cannot be categorized discretely, but rather represents a continuum from Mutualism to parasitism.
- How to retain tolerance to commensals, while pathogens are attacked?
- The range of host-microbe interactions evokes ignorance to non-harmful microbes, tolerogenic immunity to beneficial microbes (harmful if not regulated) and cytotoxic immunity to harmful microbes.

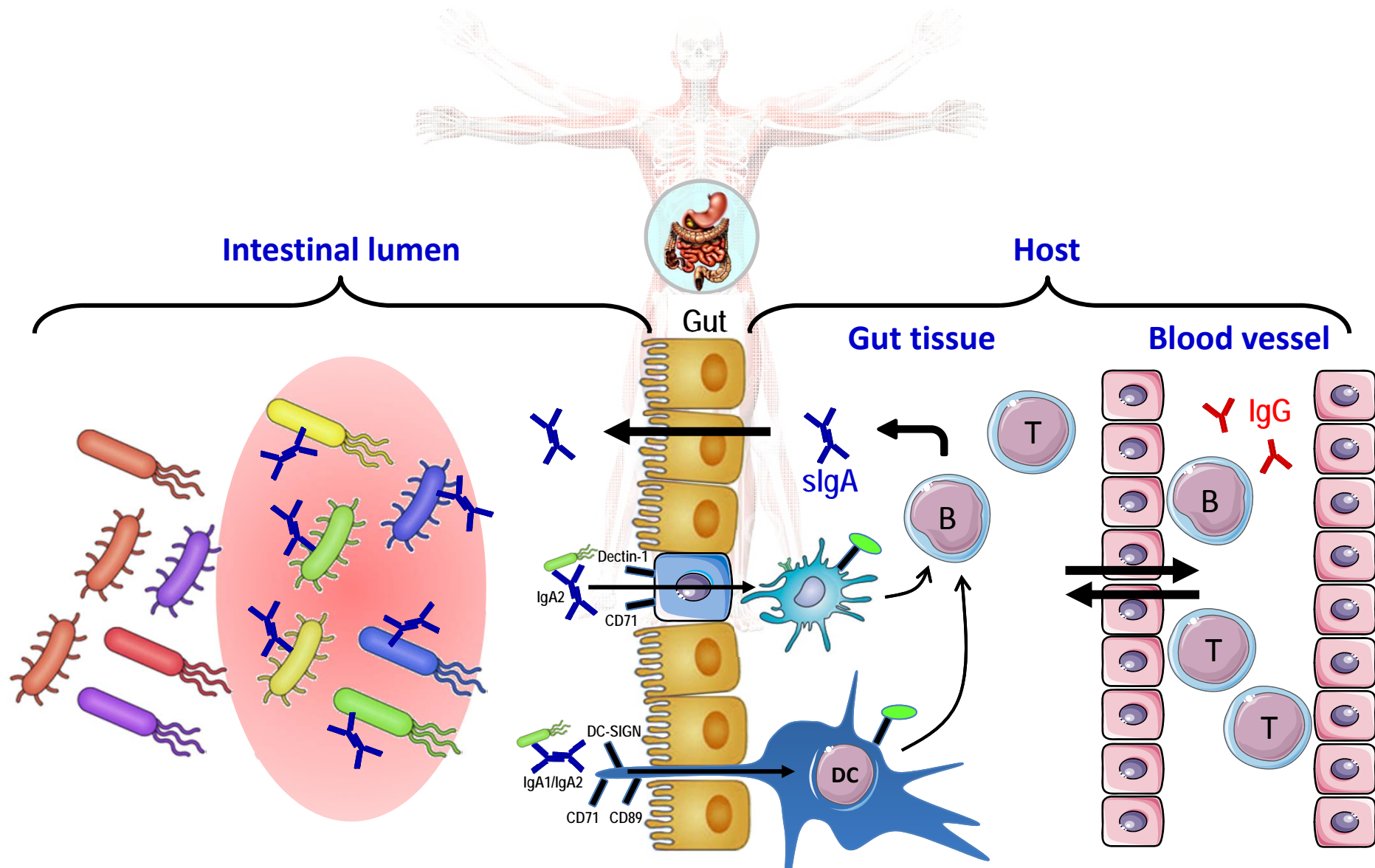
Interactions between host and gut microbiota



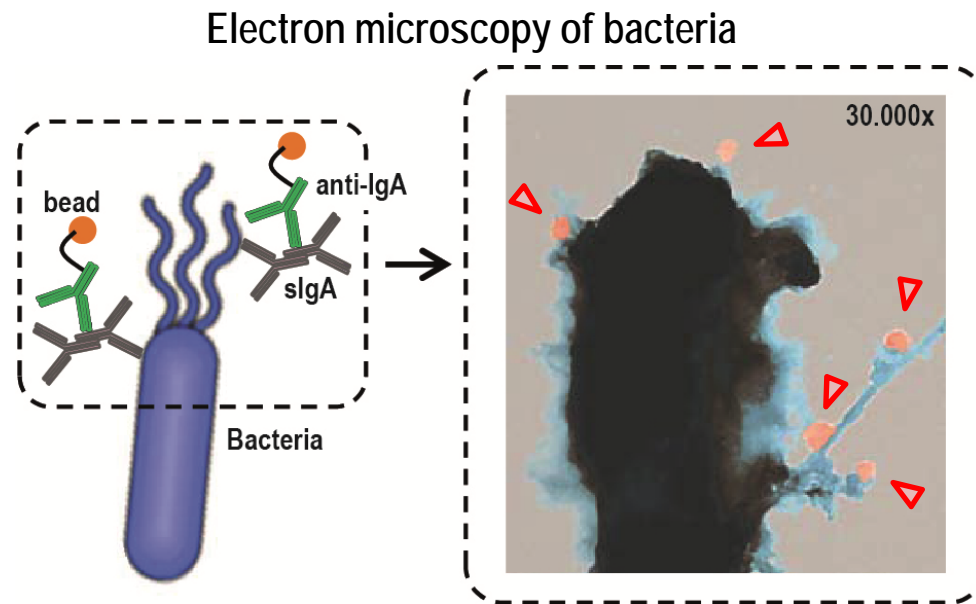
Interactions between host and gut microbiota



Interactions between host and gut microbiota

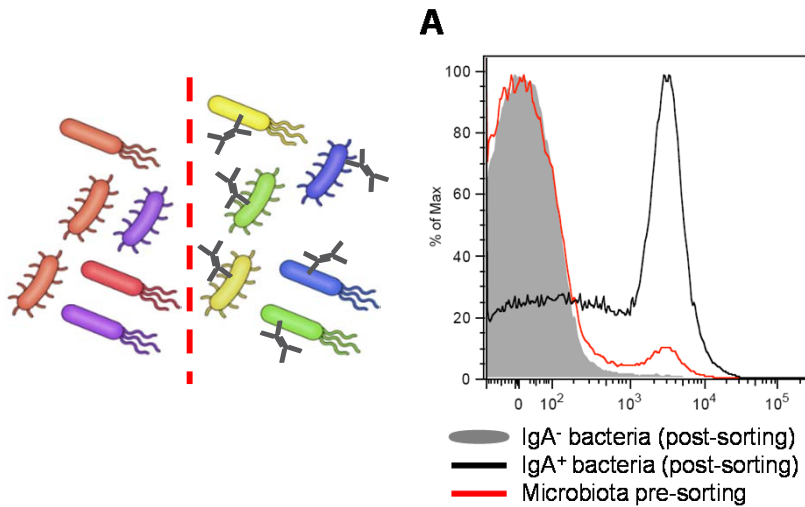


slgA opsonization of gut commensal



Gut microbiota specificity of gut Ig immunity

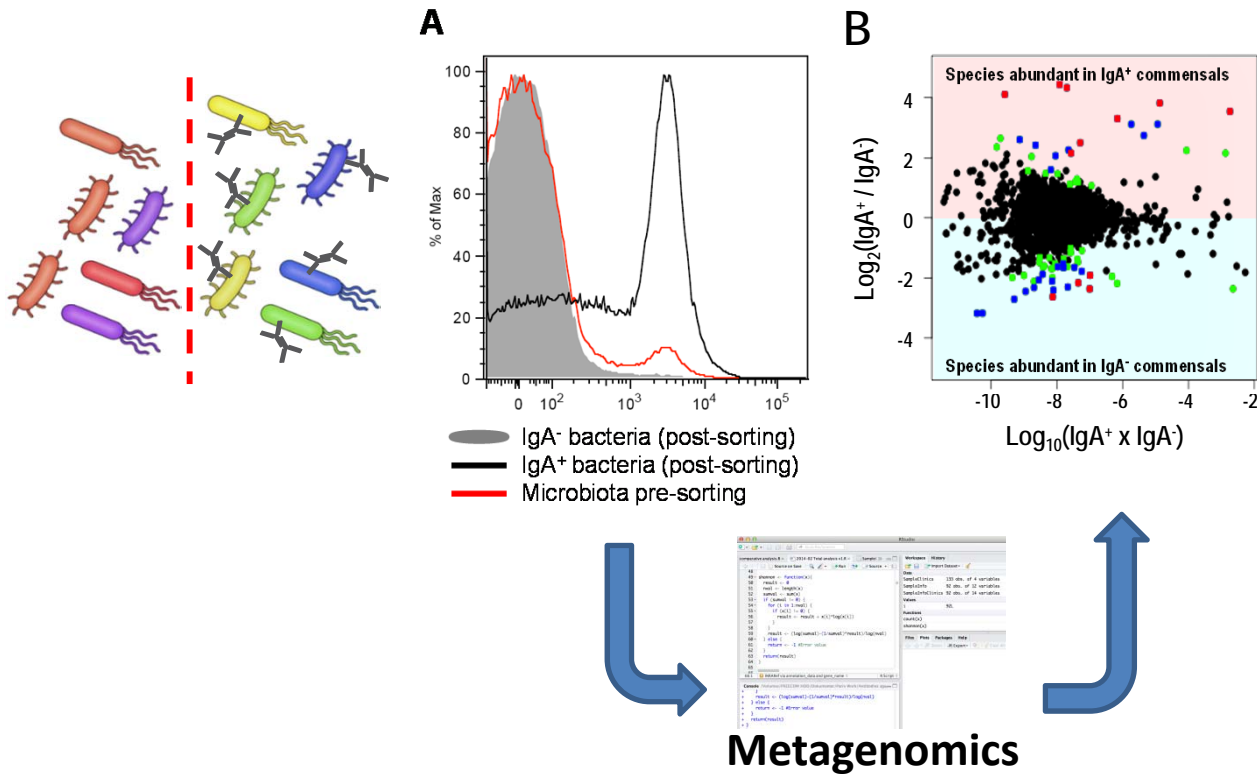
Gut microbiota sorting (IgA^+ / IgA^-)



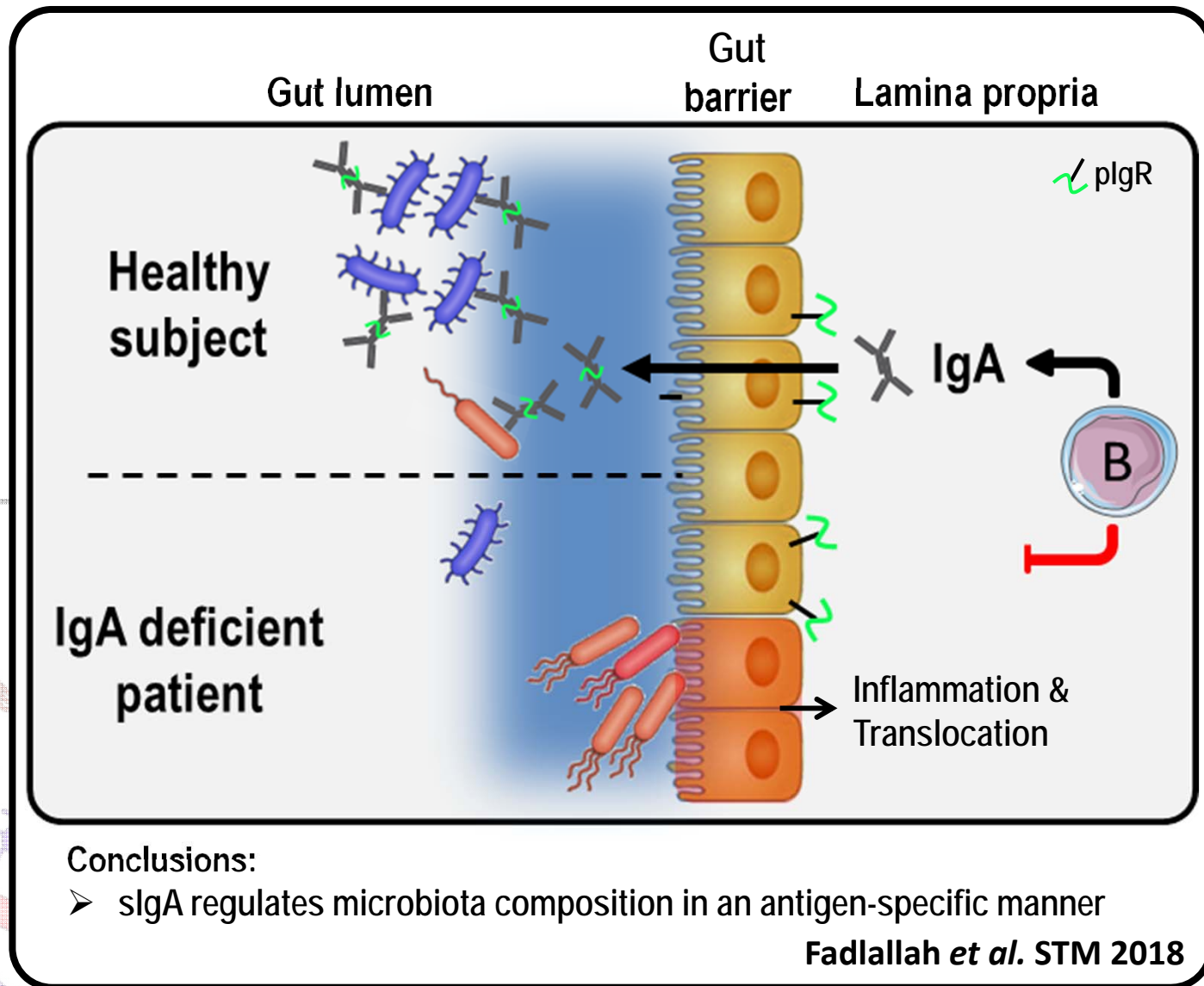
Gut microbiota specificity of gut Ig immunity

Gut microbiota sorting (IgA^+ / IgA^-)

Paired metagenomic analysis

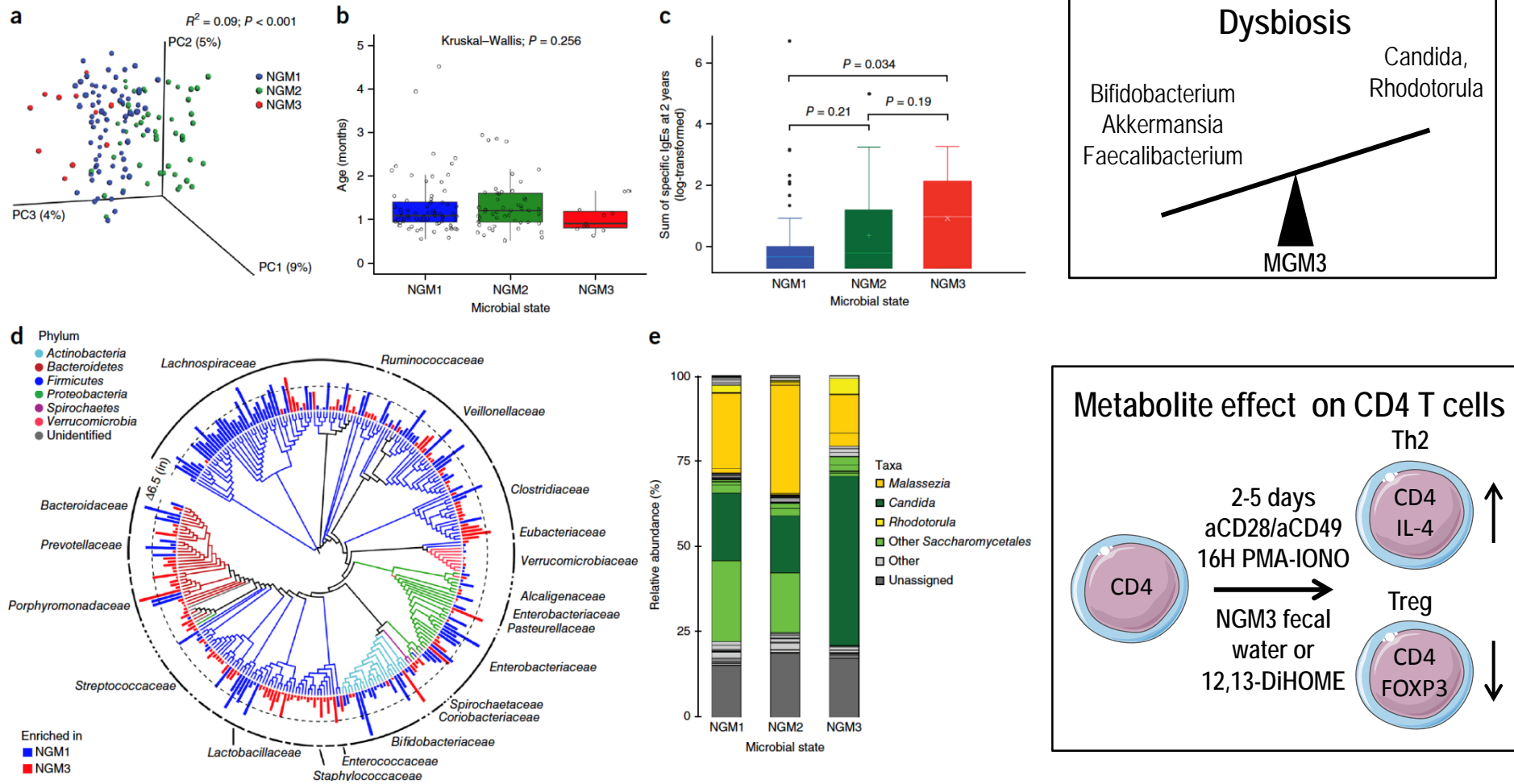


Interactions between host and gut microbiota



Examples of gut microbiota associations with allergy

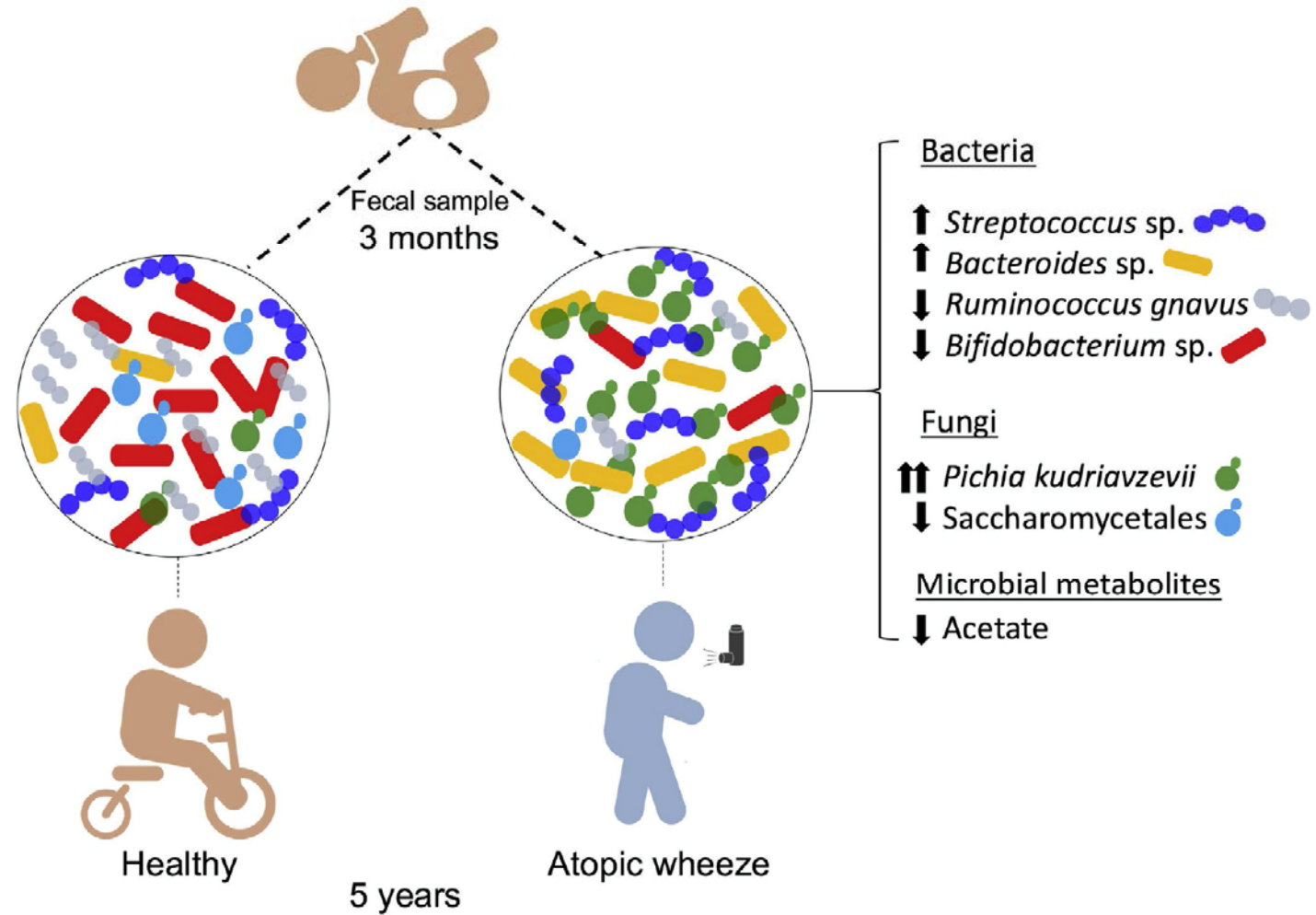
Gut Microbiota composition is associated with allergic disease



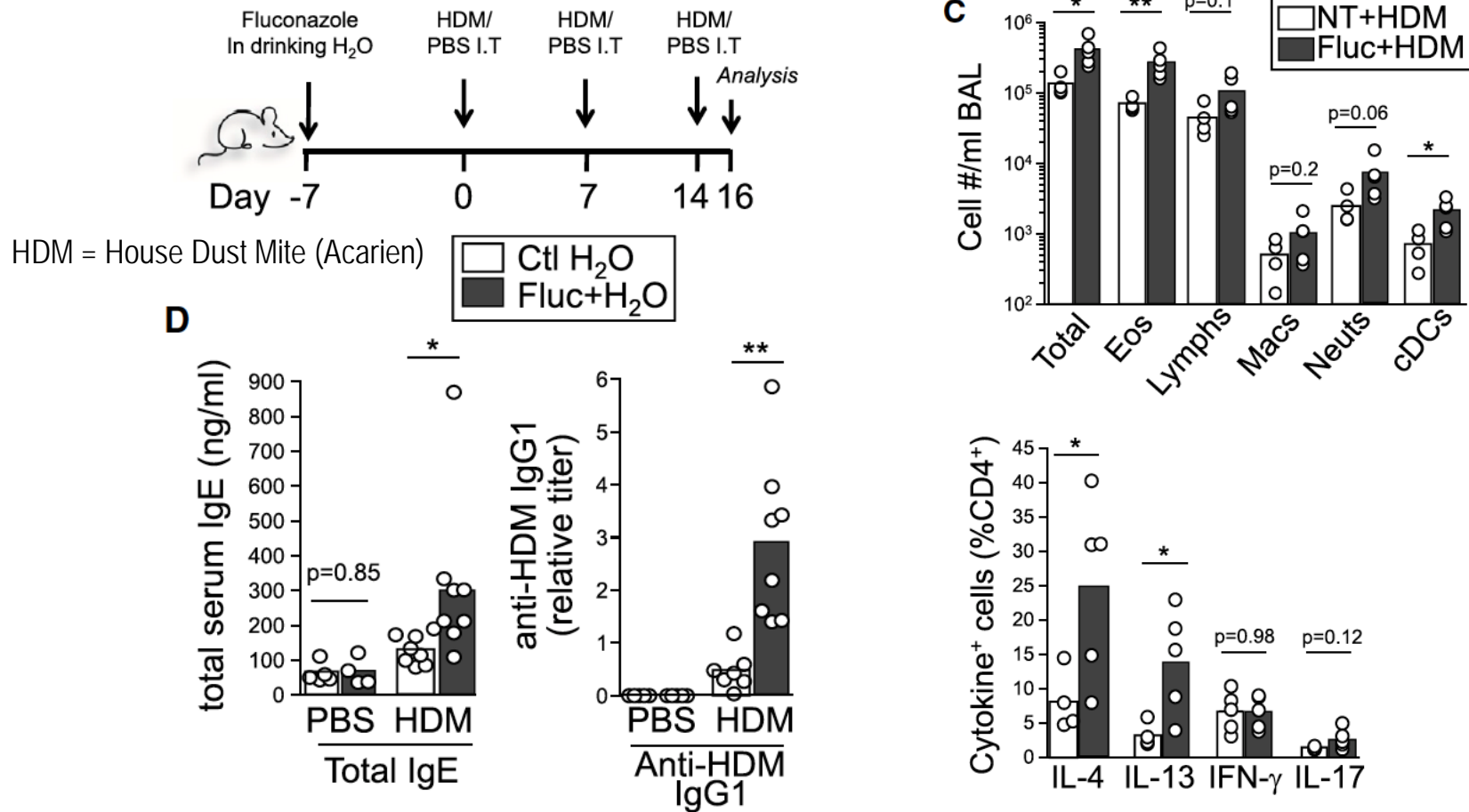
- Neonatal gut microbiota (NGM) clusters differentiate low and high risk allergic disease (loss of bacteria and gain of fungus increase risk of allergy).
- Dysbiosis alters the metabolic profile of fecal water (reduction of Tregs – increase in Th2 cells).

Bacterial and fungal dysbiosis in atopic wheeze

Dysbiosis and Atopic Wheeze in Rural Ecuador: A Case-Control Study

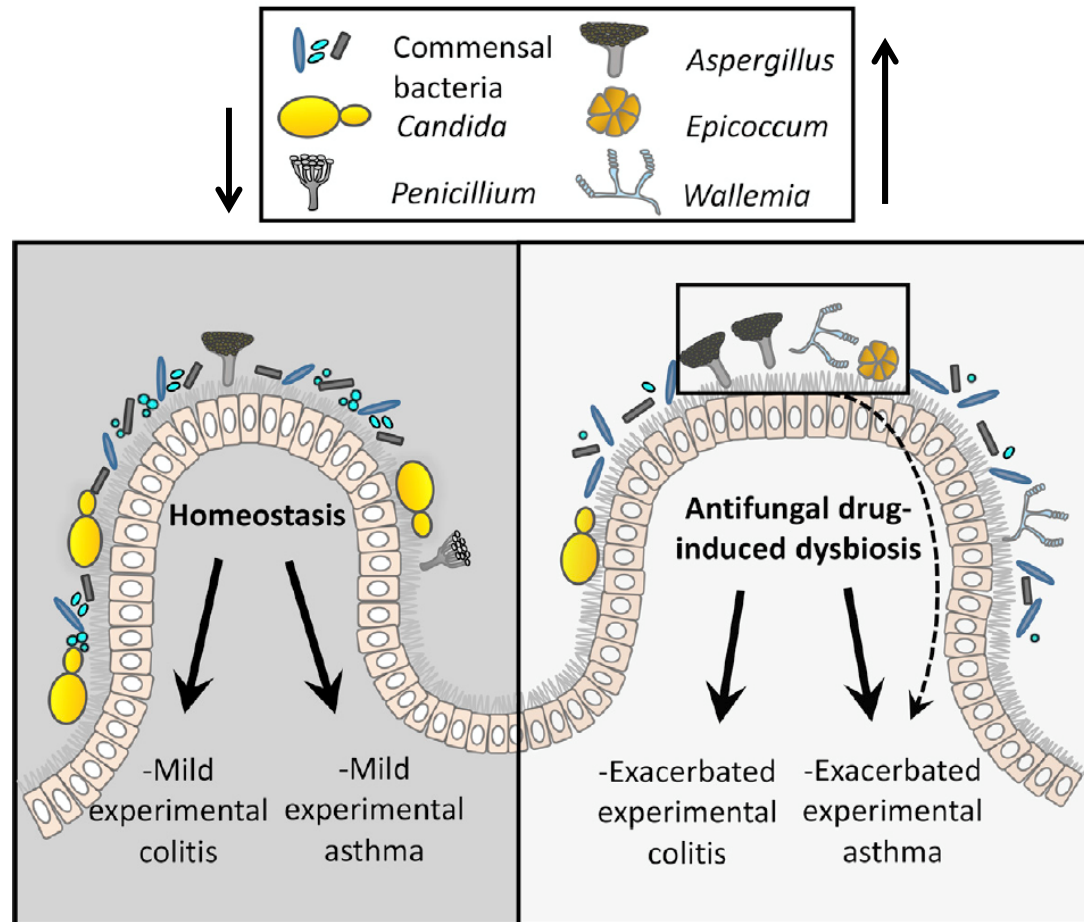


Fungal dysbiosis exacerbate atopic wheeze



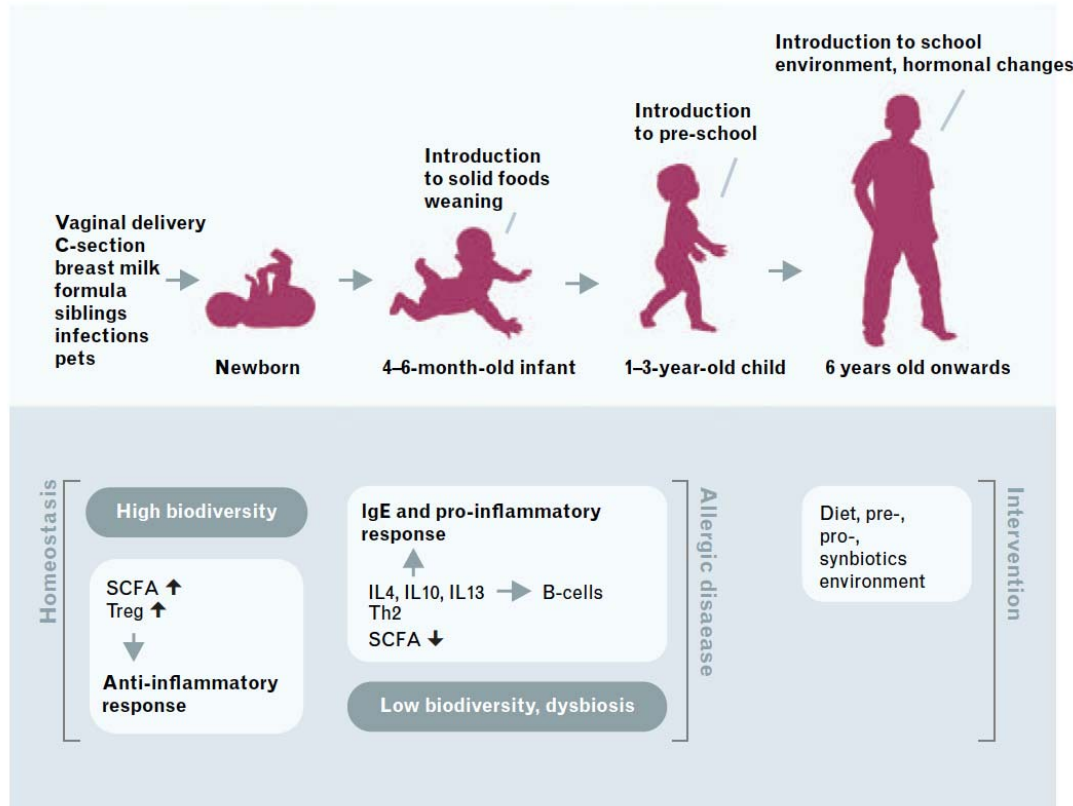
- Oral antifungals (fluconazole and amphotericin) increase the severity of allergic airway disease in mice.
- Antifungal drugs induce dysbiosis in which some fungi are reduced (*Candida* and *Penicillium*) and others expand (***Aspergillus*, *Wallemia*, and *Epicoccum spp.***)
- The three expanded fungi are sufficient to increase severity of allergic airway disease

Fungal dysbiosis exacerbate atopic wheeze



- Oral antifungals (fluconazole and amphotericin) increase the severity of allergic airway disease in mice.
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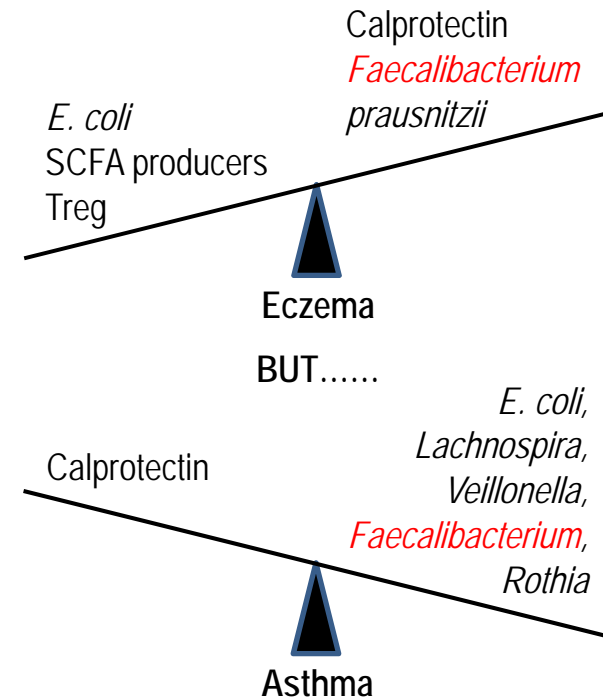
Gut microbiota and eczema



Predictive markers:

High fecal calprotectin (inflammation) and low abundance of fecal *E. coli* at 2 months predicted asthma and eczema by the age of 6 years.

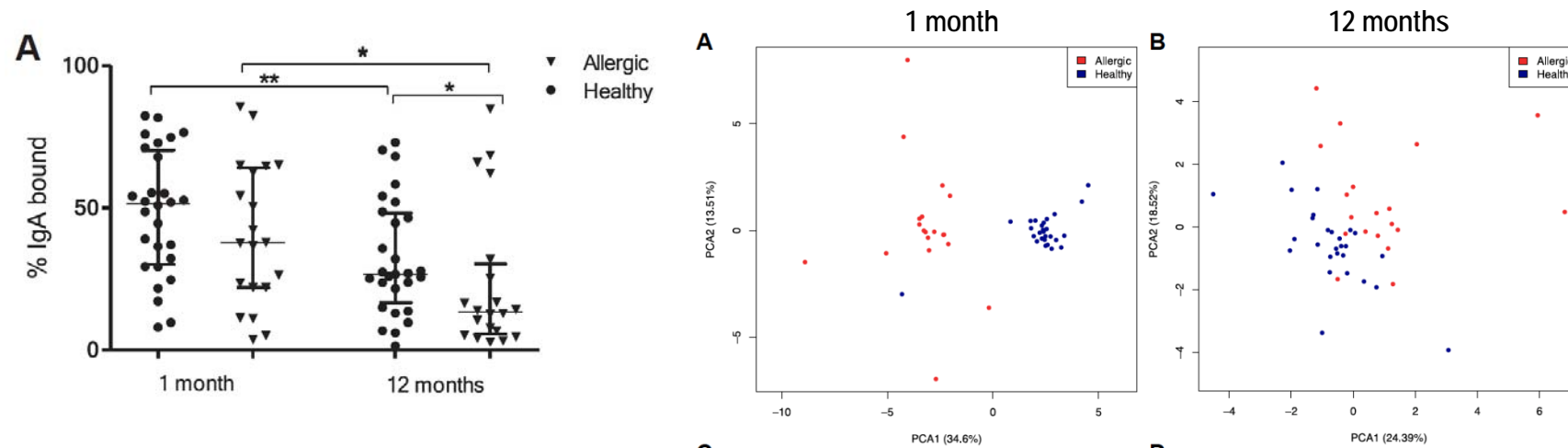
Ruminococcaceae reduced at 1 week of age in infants that developed IgE-associated eczema.



Treatment

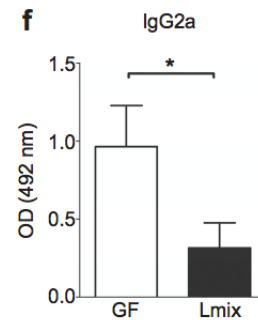
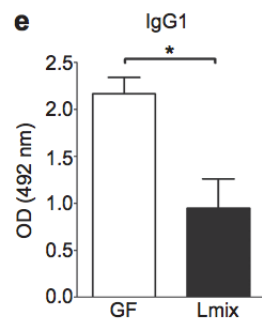
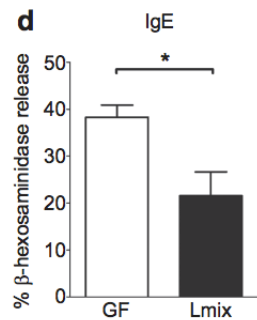
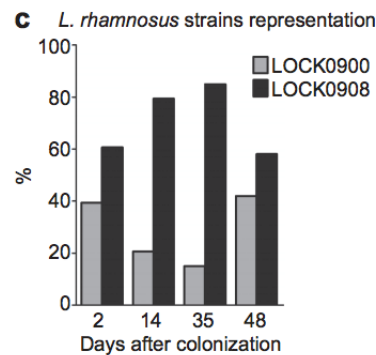
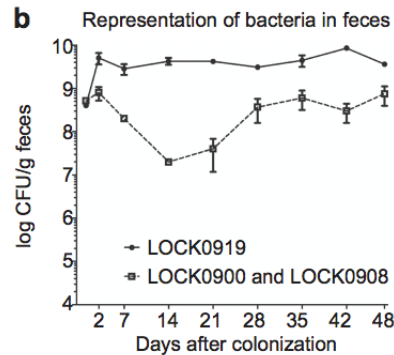
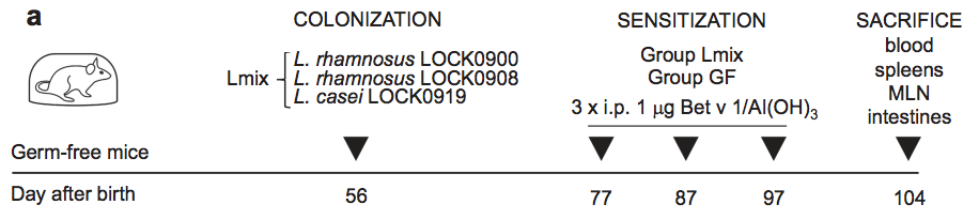
Pre- and probiotics have effect in mice, but so far not convincingly in humans.

Gut Immuno-Microbiome - allergic disease



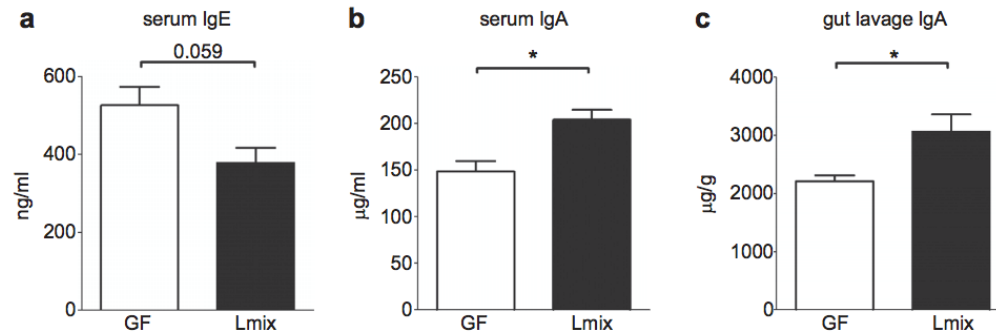
- Allergy is associated with reduced sIgA-opsionization levels of gut microbiota in 12 months-old children (tendency already at 1 month).
- Fecal sIgA levels are unaltered.
- Gut bacterial load is reduced at 12 months of age in allergic children
- A unique immuno-microbiome signature precedes allergy already at 1 month of age.
- sIgA-opsionization and immuno-microbiome signature are similar when stratified for birth route and breastfeeding (Of note, breastfeeding is assessed at 12 months – practically all Swedish children are breastfed in the first months of life)

GF vs Lactobacillus colonization - Birch pollen sensitization



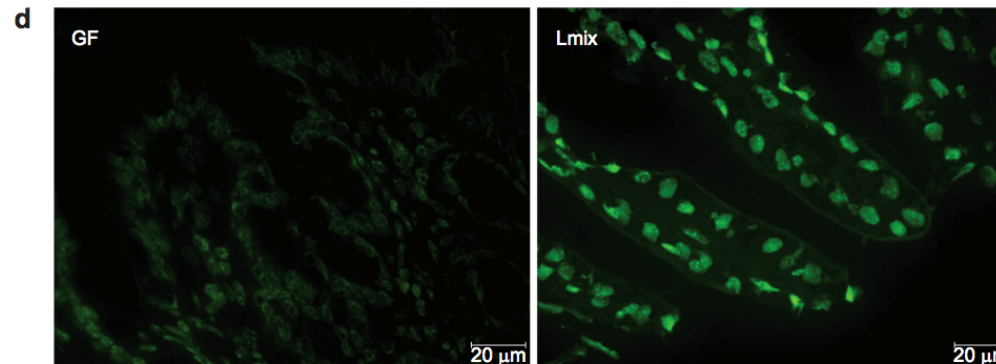
- Lactobacillus strains enhance gut barrier integrity (ZO-1; not shown) and create tolerance to birch pollen sensitization (reduced antibody responses to Bet v1)

GF vs Lactobacillus colonization - Birch pollen sensitization

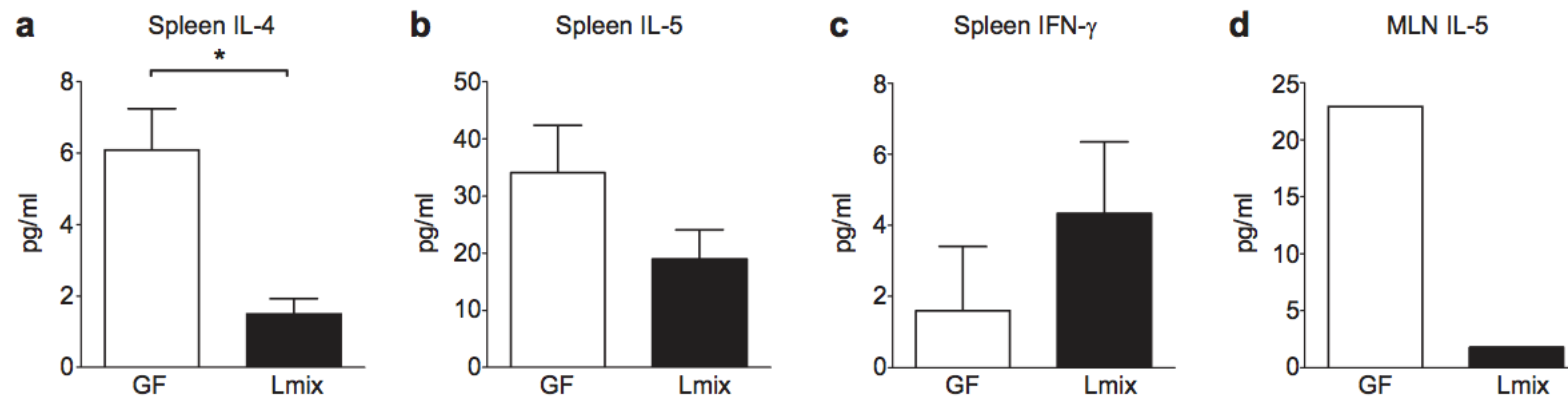


➤ Total antibody titers are shifted from IgE to IgA in lactobacillus colonized mice compared to GF.

➤ **IgA⁺ B cells (green)** are absent in the small intestine of GF mice.

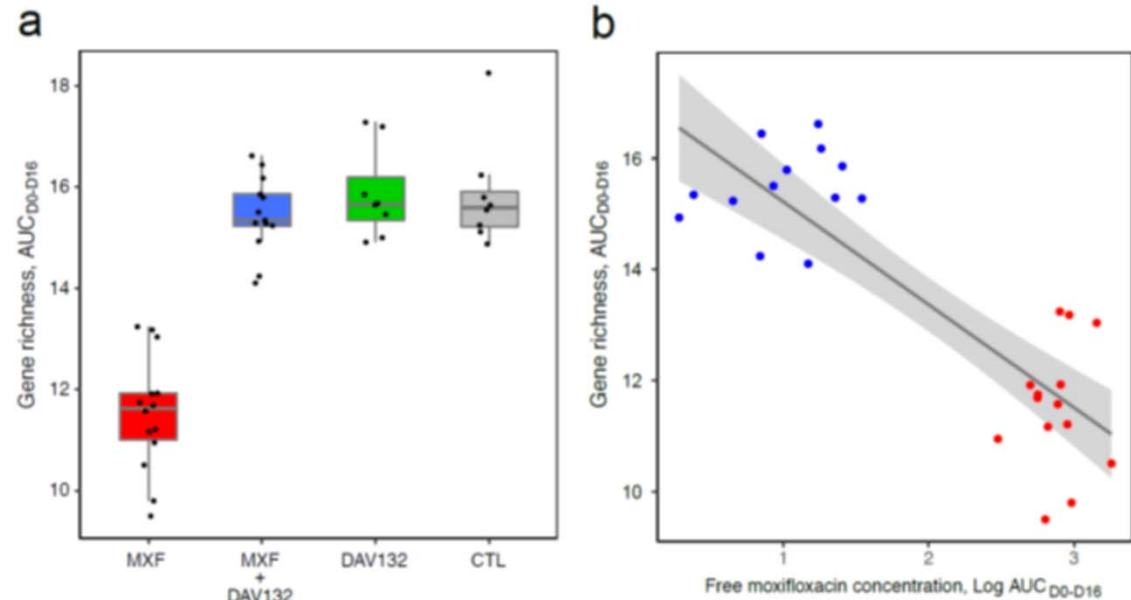
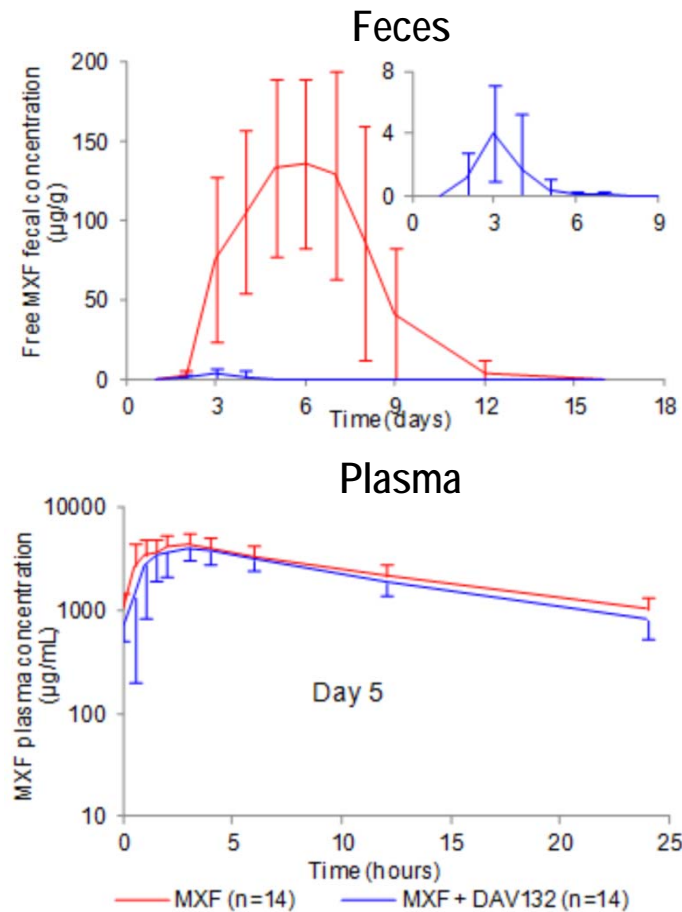


➤ Cytokine levels in the spleen shifts from Th2 to Th1 type cytokines.



Avoid intestinal effect of antibiotics

Antibiotics levels



- Antibiotics administered orally for non-intestinal infections (dental, sinus, pulmonary etc.) are taken up primarily in the stomach. Remains pass through our lower GI tract.
- Activated coal (DAV132) absorbs/inactivates 99% of antibiotics (e.g. Moxifloxacin, MXF).
- DAV132 retains normal antibiotic plasma levels, but reduces fecal levels and protects gut microbiota from antibiotic induced dysbiosis.

Take home message

- Gut microbiota influence host immunity (may skew immunity towards Th2 and IgE – to be confirmed?)
- Gut microbiota is regulated by host immunity (innate and adaptive (e.g. IgA))
- **Altered lifestyle** influence our gut microbiota composition and is temporally (but maybe not causally) associated with a rapid increase in chronic inflammatory diseases, including allergy (since 1950 forward).
- **Hygiene theory:** Reduced exposure to microbes result in a skewed host immunity, which has not been sufficiently schooled to regulate inflammatory responses.
- **Save our microbiota:** Vaginal microbiota transplantation (C-section birth), reduce antibiotics use (or use of new treatments, such as DAV132 co-therapy).
- **Save our immunity:** Probiotics (do not colonize), helminths (worms), immune therapy (allergy), promote breast feeding.